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(71) Applicant (*for all designated States except US*):
**VLAAMS INTERUNIVERSITAIR INSTITUUT
VOOR BIOTECHNOLOGIE VZW** [BE/BE]; Rijkswis-
chestraat 120, B-9052 Zwijnaarde (BE).

(72) Inventors; and

(75) Inventors/Applicants (*for US only*): **CONTRERAS,
Roland, Henry** [BE/BE]; Molenstraat 53, B-9820 Merel-
beke (BE). **CHEN, Cuiying** [BE/BE]; Lijsterstraat 10,
B-9860 Balegem (BE).

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(54) Title: METHOD TO ISOLATE GENES INVOLVED IN AGING

(57) Abstract: The present invention relates to a method to isolate genes involved in aging and/or oxidative stress, by mutation or transformation of a yeast cell, subsequent screening of the mutant or transformed cells that are affected in aging and isolation of the affected gene or genes, and the use of these genes to modulate aging and aging-associated diseases in a eukaryotic cell and/or organism.

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METHOD TO ISOLATE GENES INVOLVED IN AGING

The present invention relates to a method to isolate genes involved in aging and/or aging- associated diseases and/or oxidative stress, by mutation or transformation of a yeast cell, subsequent screening of the mutant or transformed cells that are affected in aging and isolation of the affected gene or genes, and the use of these genes to modulate aging and aging-associated diseases in a eukaryotic cell and/or organism.

Aging is a process in which all individuals of a species undergo a progressive decline in vitality leading to aging-associated diseases (AAD's) and to death. The process of aging is influenced by many factors, including metabolic capacity, stress resistance, genetic stability and gene regulation (Jazwinski, 1996). The final life span of an organism is also affected by the sum of deleterious changes and counteracting repair and maintenance mechanisms (Johnson *et al.*, 1999).

Several approaches have been followed to study aging. These include the identification of key genes and pathways important in aging, the study of genetic heritable diseases associated with aging, physiological experiment and advanced molecular biology studies of model organisms. Among these organisms, *Caenorhabditis elegans*, *Drosophila melanogaster* and the budding yeast *Saccharomyces cerevisiae* have a life span that can be influenced by single gene mutations or overexpression of a particular protein (Johnson *et al.*, 1999). Especially *S. cerevisiae* has been used as one of the model organisms to study the aging process (Gershon and Gershon, 2000). Yeast life span is defined as the number of daughter cells produced by mother cells before they stop dividing. This yeast cell divides asymmetrically, giving rise to a larger mother cell and a smaller daughter cell, leaving a circular bud scar on the mother cell's surface at the site of division. Thus, the age (counted in generations) of a mother cell can simply be determined by counting the number of bud scars on its surface. However, counting of the bud scars is labour intensive and time consuming and cannot be used as such as a screening method to isolate cells with an increased life span. Methods to isolate mutant yeasts with an increased life span have, amongst others, have been described in WO9505459 and US5874210. The latter patent describes a method to isolate a mutation which increases the number of divisions of yeast cells, comprising the labelling of the cell surface of the yeast cell with a fluorescent marker, thereby generating fluorescent yeast cells, culturing the yeast cells under conditions for

growth of yeast cells for a period of time greater than the chronological life span of the strain, selecting the fluorescent cells by fluorescence-activated cell sorting and replating the fluorescent yeast cells. However, although this method may indeed give an enrichment of strains that survive longer, there is no direct selection for strains with an increased number of divisions, and non-dividing or slower dividing cells that also survive may be selected too.

In this invention, we disclose a method for specific isolation of old yeast mother cells, with an increased number of divisions by staining the bud scar chitin with fluorescein isothiocyanate (FITC)-wheat germ agglutinin (WGA) lectin and sorting by a FACS apparatus, after initial enrichment of the mother cells through magnetic-based sorting. The process is presented in Figure 1. Said method can be used to isolate genes or mutations involved in aging.

Much attention has been focussed on the hypothesis that oxidative damage plays an important role in aging (Shan *et al.*, 2001; Hamilton *et al.*, 2001) and there is a generally accepted relation between oxidative stress and aging (Tanaka *et al.*, 2001). Moreover, mutations in genes related to protection against oxidative stress have a clear influence on life span, both in *S. cerevisiae* and *Caenorhabditis elegans* (Laun *et al.*, 2001; Ishii, 2001). This makes that the method, proposed here, is also suitable as an indirect selection for genes involved in oxidative stress. This is especially useful in cases where screening of libraries in an endogenous system is difficult or impossible, such as the screening of mammalian or plant libraries. Screening of such libraries may lead to new genes involved in protection against oxidative stress in general, but also, in case of mammalian cells, to genes involved in AAD's and/or diseases caused by oxidative stress, especially neurodegenerative diseases such as Alzheimer's disease, Parkinson's disease and Huntington's disease (Calabrese *et al.*, 2001)

A frequently practiced strategy in searching genes responsible for aging is by selecting survivals after exposure cells to stresses. Then a question constantly remaining is whether the genes picked up are in response to the stress treatment rather than involved in aging, because of the complexity of the process. The invention described here, however, provides an alternative that allows direct hunting of genes with potential anti-aging functions from various libraries or library combinations of eukaryotic organisms. Yeast lines are selected in a more natural condition, and also with advantages of high throughput, high efficiency, and short

time investment. Obviously, this invention has a great potential for rational drug design and development of therapies and prevention in the field of age-related diseases.

- 5 It is a first aspect of the invention to provide a method to screen genes involved in aging and/or AAD's and/or oxidative stress, comprising a) mutation or transformation of a yeast cell b) cultivation of said cell c) enrichment of the population for mother cells d) labelling said mother cells with a WGA- based label and e) isolation of the highly labelled cells.
- 10 To obtain a sufficient distinction between old cells and young cells, it is essential to use a marking of the bud scars that is sufficiently linear with the number of scars, and is not or only weakly interacting with other cell wand compounds. Surprisingly we found that WGA can bind with the chitin in the bud scar, without major interference with other cell compounds, so that the amount of WGA bound is a
- 15 reliable measurement of the number of bud scars. The WGA bound is then measured using a WGA-based label. A WGA-based label, as used here, may be any kind of label that allows quantifying the amount of WGA bound to the cell and may be, as a non-limiting example, WGA coupled to a stain, or a detectable antibody that binds to WGA. Detectable antibodies are known to the person skilled
- 20 in the art and may be, as a non-limiting example, rabbit antibodies that can be detected by a labelled anti-rabbit antibody. The labelling of mother cells with a WGA based label may be a one step process, whereby labelled WGA is bound to the cell, or a two step process, whereby in a first step, WGA is bound to the bud scars, and in a second step, the bound WGA is labelled. A preferred embodiment is a method
- 25 according to the invention, whereby said WGA based label is FITC labelled WGA. Preferably, said isolation of highly stained cells is based on FACS sorting. Methods for the enrichment of the population of mother cells are known to the person skilled in the art and may be based on, as a non-limiting example, staining of the cell wall of the cells at a certain point in the growth phase, followed by continuation of the
- 30 culturing and sorting of the stained cells. Alternatively, the cells may be antibody labelled. Preferably, said enrichment of the population of mother cells is a magnetic-based sorting. Instead of being based on a global cell wall labelling as described above, the enrichment of the population of mother cells may be based on the labelling of a fraction of the mother cells, such as a bud scar based labelling. In

fact, the enrichment of the mother cells may be carried out by a first WGA based labelling and sorting, whereby the enriched mother cells are subjected to a second WGA based labelling and sorting. The labelling method in the first and second round may be different.

5 Methods to mutate yeasts are known to the person skilled in the art and include, but are not limited to chemical and physical mutagenesis, such as ethyl methane sulphonate (EMS) treatment, or UV treatment. Methods to transform yeast are also known to the person skilled in the art and include, but are not limited to protoplast transformation, lithium acetate based transformation and electroporation. The yeast
10 transformation may be carried with one or more nucleic acids, up to a complete library. The nucleic acid used is not necessarily yeast nucleic acid, but may be from any origin, as long as it is functionally expressed in yeast. Preferred examples of nucleic acids are mammalian nucleic acids, such as human nucleic acid, and plant nucleic acid, whereby said nucleic acids are cloned in a yeast expression vector.
15 Preferably, the yeast is transformed with an expression library. The nucleic acid that is transcribed into mRNA does not necessarily be translated into protein, but may exert its effect as antisense RNA. Indeed, it is an additional advantage of the method that it can detect in one screening experiment both the effect of overexpression of a protein, as well as the effect of downregulation of a protein by
20 blocking the translation of an endogenous messenger by a homologous antisense RNA, resulting from the expression library.

Another aspect of the invention is a gene or functional gene fragment isolated with the method, according to the invention. Said functional fragment may encode for a polypeptide, that directly affects aging and/or an AAD and/or oxidative stress, or it
25 may be transcribed into antisense RNA, which affect aging and/or an AAD and/or oxidative stress by silencing an endogenous gene. Preferably, said gene or functional gene fragment is selected from the nucleic acid listed in table 2. More preferably, said gene or functional gene fragment comprises a sequence as represented in SEQ ID N° 1, 3, 5, 7, 8, 9, 11, 13, 15, 16, 17, 19, 21, 22, 23, 24, 25,
30 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52 or 53. Even more preferably, said gene or gene fragment is essentially consisting of a sequence as represented in SEQ ID N° 1, 3, 5, 7, 8, 9, 11, 13, 15, 16, 17, 19, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52 or 53. Even more preferably,

said gene or functional gene fragment is consisting of a sequence as represented in SEQ ID N° 1, 3, 5, 7, 8, 9, 11, 13, 15, 16, 17, 19, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52 or 53. A preferred embodiment is a gene fragment, isolated with the method, essentially consisting of SEQ ID N° 11, preferably consisting of SEQ ID N° 11.

Another preferred embodiment is a gene fragment, isolated with the method, essentially consisting of SEQ ID N° 16, preferably consisting of SEQ ID N° 16.

Still another aspect of the invention of the use of a gene or functional gene fragment isolated with the method according to the invention to modulate aging and/or to modulate the development of AAD's and/or to protect against oxidative stress. Preferably, said modulation is an inhibition of aging. Preferably, said gene or gene fragment is selected from the nucleic acids listed in table 2. More preferably, said gene or gene fragment comprises a sequence as represented in SEQ ID N° 1, 3, 5, 7, 8, 9, 11, 13, 15, 16, 17, 19, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52 or 53. Even more preferably, said gene or gene fragment is essentially consisting of a sequence as represented in SEQ ID N° 1, 3, 5, 7, 8, 9, 11, 13, 15, 16, 17, 19, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52 or 53. Even more preferably, said gene or gene fragment is consisting of a sequence as represented in SEQ ID N° 1, 3, 5, 7, 8, 9, 11, 13, 15, 16, 17, 19, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52 or 53. A preferred embodiment is the use of a functional gene fragment, essentially consisting of SEQ ID N° 11, preferably consisting of SEQ ID N° 11. Another preferred embodiment is the use of a gene fragment, isolated with the method, essentially consisting of SEQ ID N° 16, preferably consisting of SEQ ID N° 16.

Another aspect of the invention is a polypeptide, encoded by a gene or functional gene fragment isolated with a method according to the invention. Preferably, said modulation is an inhibition of aging and/or inhibition of the development of an AAD. Preferably, said polypeptide is encoded by a nucleic acid listed in table 2. More preferably, said polypeptide is encoded by a nucleic acid comprising SEQ ID N° 1, 3, 5, 7, 8, 9, 11, 13, 15, 16, 17, 19, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52 or 53. Even more preferably, said polypeptide is encoded by a nucleic acid essentially consisting of

SEQ ID N° 1, 3, 5, 7, 8, 9, 11, 13, 15, 16, 17, 19, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52 or 53. Even more preferably, said polypeptide is encoded by a nucleic acid consisting of SEQ ID N° 1, 3, 5, 7, 8, 9, 11, 13, 15, 16, 17, 19, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52 or 53. Even more preferably, said polypeptide comprises SEQ ID N° 2, 4, 6, 10, 12, 14, 18, or 20. Even more preferably, said polypeptide is essentially consisting of SEQ ID N° 2, 4, 6, 10, 12, 14, 18 or 20. Even more preferably, said polypeptide is consisting of SEQ ID N° 2, 4, 6, 10, 12, 14, 18 or 20. A preferred embodiment is a polypeptide, essentially consisting of SEQ ID N° 12, preferably consisting of SEQ ID N° 12. Still another preferred embodiment is a polypeptide encoded by a nucleic acid essentially consisting of SEQ ID N° 16, preferably consisting of SEQ ID N° 16

Still another aspect of the invention is the use of a polypeptide, encoded by a gene or functional gene fragment, isolated with a method according to the invention, to modulate aging and/or to modulate the development of an AAD and/or to protect against oxidative stress. Preferably said modulation is an inhibition of aging and/or inhibitor of the development of an AAD. Preferably, said polypeptide is encoded by a nucleic acid selected from the nucleic acids listed in table 2. More preferably, said polypeptide is encoded by a nucleic acid comprising SEQ ID N° 1, 3, 5, 7, 8, 9, 11, 13, 15, 16, 17, 19, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52 or 53. More preferably, said polypeptide comprises SEQ ID N° 2, 4, 6, 10, 12, 14, 18 or 20. Even more preferably, said polypeptide is essentially consisting of SEQ ID N° 2, 4, 6, 10, 12, 14, 18 or 20.

Most preferably, said polypeptide is consisting of SEQ ID N° 2, 4, 6, 10, 12, 14, 18 or 20. A preferred embodiment is the use of a polypeptide, essentially consisting of SEQ ID N° 12, preferably consisting of SEQ ID N° 12, to modulate aging and/or to modulate the development the development of an AAD. Preferably, said modulation is an inhibition of aging and/or an inhibition of the development the development of an AAD. Still another preferred embodiment is the use of a polypeptide, encoded by a nucleic acid comprising SEQ ID N° 16, preferably essentially consisting of SEQ ID N° 16, more preferably consisting of SEQ ID N° 16, to modulate aging and/or to modulate the development the development of an AAD.

Still another aspect of the invention is the use of an antisense RNA encoded by a gene or a functional gene fragment, isolated with a method according to the invention, to modulate aging and/or to modulate the development the development of an AAD. In such an application, the gene or functional gene fragment is operationally linked to a promoter, in such a way that an antisense RNA, complementary to the mRNA encoding the polypeptide normally encoded by said gene or gene fragment, is transcribed. Preferably, said gene or functional gene fragment encoding the antisense RNA comprises SEQ ID N° 7, 8 or 15. Even more preferably, said modulation of aging is an inhibition of aging and/or an inhibition of the development the development of an AAD.

Definitions

Gene as used here refers to a region of DNA that is transcribed into RNA, and subsequently preferentially, but not necessarily, translated into a polypeptide. The term is not limited to the coding sequence. The term refers to any nucleic acid comprising said region, with or without the exon sequences, and includes, but is not limited to genomic DNA, cDNA and messenger RNA. As, on the base of these sequences, it is evident for the person skilled in the art to isolate the promoter region, the term gene may include the promoter region when it refers to genomic DNA.

Nucleic acid as used herein refers to a polymeric form of nucleotides of any length, either ribonucleotides or deoxyribonucleotides. This term refers only to the primary structure of the molecule. Thus, this term includes double- and single-stranded DNA, and RNA. It also includes known types of modifications, for example, methylation, "caps" substitution of one or more of the naturally occurring nucleotides with an analog.

Functional fragment of a gene involved in aging is every fragment that, when tested with the method according to the invention, still gives a positive response. Typically, functional fragment are fragments that have deletions in the 5' and/or 3' untranslated regions. Alternatively, the functional fragment may be an antisense fragment, encoding an RNA that is silencing an endogenous gene, or functions as RNAi. As the coding sequence on its own is also considered as a functional fragment, as it is evident for the person skilled in the art that it may be functional when it is placed between suitable heterologous 5' and 3' untranslated sequences.

Polypeptide refers to a polymer of amino acids and does not refer to a specific length of the molecule. This term also includes post-translational modifications of the polypeptide, such as glycosylation, phosphorylation and acetylation.

5 *Aging* as used here includes all forms of aging, particularly also aging-associated diseases (AAD's). AAD's are known to the person skilled in the art and include, but are not limited to arteriosclerosis, Parkinson's disease and Alzheimer's disease.

Brief description of the figures

10 Figure 1. Scheme of the bud scar sorting (BSS) system for yeast M-cells. The BSS system contains two major steps. The first step at the left side of the figure, magnetic sorting of biotinylated M-cells and re-growth of sorted M-cells to desired generations when needed. The second step at the right side of the figure, WGA staining of bud scars and sorting of longer life M-cells according to bud scar staining.

15 Figure 2. Flow cytometric assay of yeast cells labelled with WGA-FITC and streptavidin-PE.

Yeast cells (M-cell) are grown for 5 to 6 generations (G5-6) after biotin labelling, sorted via MACS, and then simultaneously labelled with WGA-FITC and streptavidin-PE. A: shows a clear separation of the PE red-fluorescent mother cells (gated M-cell) from the non-PE fluorescent daughter cells (gated D-cell). B: hardly detects the PE fluorescent signal in the depleted daughter cells. C and D: the layout of FSC versus SSC, the gated M-cells mainly appeared at higher FSC/SSC values representing a large cell size population (C) compared to a small cell size population of D-cells at lower FSC/SSC values (D). E and F: the M-cell population gives strong WGA-FITC staining (E) than the D-cell population (F).

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Figure 3. Bud scar staining of yeast cells. INVSc-1 cells (M-cells) were biotinylated and cultured in SD medium. M-cells at G5-6 were magnetically sorted. Staining of bud scars with WGA-FITC was revealed with a Zeiss LSM410 confocal microscope.

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Figure 4. Screen of a human cDNA library via FACS.

A cDNA library from HepG2 hepatoma cells was transformed into the yeast strain INVSc-1 (pEX2) (See Materials and Methods). The transformed yeast population was first labeled with biotin and then cultured in S-glycerol medium. The initial biotinylated M-cells of approximately G14 (14 generations) were obtained by running

35

two magnetic sorting and regrowth cycles, and were then double labelled with WGA-FITC and streptavidin-PE. The older mother cells were gated according to PE staining and big cell size which represented as high FSC (A). Flow sorted older mother cells (gate Old-M) show a strong WGA-FITC signal (B).

5

Figure 5. Flow cytometric dead cell assay using PI staining.

Flow cytometric analysis of cell death using PI staining was performed in a ferritin L chain clone (pEX2-FL) and its parent line of INVSc-1 (pEX2). Yeast cells were grown up to 6 generations. The gate R1 was set around PI-positive cells that cover the dead cells, the gate R2 around the PE-positive cells that represents the M-cell population and the gate R3 around the D-cell population. In the panel A, it shows 16,3% dead cells for the ferritin L chain clone. In panel B, a 33% dead cell was observed in the control line.

10

Figure 6. Resistance of ferritin containing yeast to H₂O₂ (1mM) stress.

Cells transformed with the plasmids as indicated were exponentially grown at 30° C to an OD₆₀₀ of approximately 0.5. Cells were treated with 1mM H₂O₂ during various times. Samples were diluted and plated on YPD solid media to monitor cell viability. Cl2-ferritin indicates the cell line containing the ferritin-fragment expression vector of pGAL10-FL. Its parent line transformed with the empty vector of pSCGAL10-SN was used as control.

15

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Figure 7. Life span of *C. elegans* carrying the human Ferritin Light Chain (*FTL*) gene. Animals were injected with a L4759 plasmid containing human *FTL* gene. Controls were injected with empty plasmids. pRF4 containing the dominant phenotypic marker *rol-6(su1006)* was coinjected in both cases. Results are cumulative from four independent experiments with more than 25 animals per trial. Life-span is defined as the day when the first transformed larvae hatched until their death. Animals carrying copies of the human *FTL* gene lived significantly longer (13.54 ± 0.269 days) than controls (12.50 ± 0.266 days).

25

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Figure 8. Study of the aging phenotype of yeast $\Delta fob1$ strain by the mixed-growth system. A mixture of $\Delta fob1$ strain and parent BY4742, were biotinylated and grown in SD medium as described in example 7. G20 (the point after 20 generations) was

obtained by running three cycles of magnetic sorting and regrowth. The results show an increased frequency of $\Delta fob1$ cycling M-cells at G20, illustrating a longer life span.

Figure 9. Comparison of the viability of *FTL* strain with its parents.

5 The initial mixture of M-cells (*FTL* and INVSc-1) was biotinylated and grown in minimal SD and S-glycerol media as described in materials and methods. The ratio of viable M-cells in the mixture at different ages was determined by plating. Data for cells grown in the *FTL* gene inducing S-glycerol medium, are presented at the right side of the figure, while data for the control are shown on the left side, indicating that
10 the difference in aging is clearly due to the ferritin expression. In a separate experiment, doubling times of both strains were carefully tested and found to be equal.

Figure 10. Ferritin L prevents fast aging in presence of iron in yeast as tested by
15 micromanipulator experiment

Life spans of human partial ferritin and full ferritin transformed in strain BY4741. S-raffinose was used as carbon source for inducing expression of ferritin. An excess of iron was added in the medium with 500 μ M FAC and 80 μ M ferrichrome. At least 60 cells were included in each of three life span assays. Both partial and full ferritin had
20 a longer average life span (17.85 G and 15.58G) than the control (12.19).

Examples

Materials and methods to the examples

25 *Strains and Media*

The following *S. cerevisiae* strains were used: INVSc-1 (Invitrogen, San Diego, CA); BY4741 and BY4742 (Euroscarf, Frankfurt, Germany) as well as the BY4742-derived $\Delta fob1$ strain (Euroscarf; accession No. Y14044). Strains were grown at 30°C in rich YPD medium (2% dextrose, 2% bactopectone and 1% yeast extract) or minimal SD
30 medium (0.67% yeast nitrogen base without amino acids, 2% dextrose and 0.077% complete supplement mixture - uracil). The INVSc-1 and BY4741 strains used for library screening were grown in S-glycerol, S-galactose or S-raffinose media, where dextrose is replaced with 3% glycerol, 2% galactose or 2% raffinose, respectively. S-glycerol was used to induce expression of genes cloned in pEX2, whereas S-

galactose was used to induce expression of genes cloned in pSCGAL10-SN. Media were solidified with 2% agar.

Cloning and overexpression of a human cDNA library

5 To recover mRNA from various responses, a pool of equal proportions of human HEPG2 cells, subjected to different treatments, was used for library construction. These treatments included heat shock for 1.5 h at 42.5°C, 1 mM dithiothreitol, 100 U/ml interleukin-6 and 10^{-7} M dexamethasone. Construction of cDNA libraries was carried out essentially as described previously (Declercq et al. 2000). cDNA was
10 cloned at the site of SfiI/NotI in the vectors pEX2 (BCCM/LMBP Plasmid Collection, Ghent University, Belgium; accession No. 2890) and pSCGAL10-SN (BCCM/LMBP Plasmid Collection, accession No. 2471). cDNA expression is driven by the cytochrome c promoter in pEX2 and by the GAL10 promoter in pSCGAL10-SN. Yeast strain INVSc-1 was used as the host for pEX2 library transformation. The pSCGAL10-SN library was transformed to the BY4741 strain. Transformations were performed as
15 described previously (Gietz and Woods, 2001). Approximately 3.5×10^5 colonies from each transformation were produced.

Magnetic sorter based preparation of yeast mother cells (M-cell)

20 Cells were cultured at 30°C in liquid medium, such as minimal SD medium or in the specific induction medium, to OD₆₀₀ of 0.7-1 and were collected by centrifugation. All cells harvested were used as M-cells. The biotin labelling of M-cells was carried out essentially as described previously (Smeal et al., 1996). Before labelling, M-cells were washed twice with cold phosphate-buffered saline (PBS; pH 8.0), resuspended in PBS
25 to a concentration of 2.5×10^7 cells/ml and then incubated with 0.1 mg/ml Sulfo-NHS-LC-Biotin (Pierce Chemical Company, Rockford, IL) for 30 min at room temperature under gentle shaking. The free biotin reagent was removed by two washings with PBS. Biotinylated M-cells were grown in liquid medium for a desired number of generations (up to G7 in our conditions; culture was not allowed to exceed OD₆₀₀ = 1).

30 The separation of mother cells from the daughter cells they produced was carried out via magnetic cell sorting. This was realized by coupling the biotinylated mother cells to magnet beads by incubating 10^7 mother cells with 80 µl of Anti-Biotin MicroBead (Miltenyi Biotec, Germany) in 1ml PBS pH 7.2 for 1hour at 4°C. Unbound

beads were removed by washing twice with PBS. M-cells were isolated with a magnetic sorter according to the supplier's protocol (Miltenyi Biotec). When needed, these sorted M-cells can be further grown in liquid medium for additional generations and isolated again by the magnetic sorting system.

- 5 The purity of sorted mother cells was determined on the basis of streptavidin binding. About 10^7 biotinylated cells were stained with 3 μ g streptavidin-conjugated R-phycoerthrin (PE) (Molecular Probes) in 1 ml of PBS pH 7.2 for 1 hour at room temperature in total darkness. Then cells were washed twice with PBS and suspended in 2 ml of PBS pH 7.2. The yeast cells with more bud scars were
10 recognised as a high intensity of FITC signals.

WGA-based bud scar staining

- The bud scars of yeast cells were stained with fluorescein isothiocyanate (FITC)-labelled WGA lectin (Sigma). The staining was carried out by adding 10^7 yeast cells
15 together with 12 μ g WGA-FITC in 1 ml of PBS pH 7.2 for 1.5 hours at room temperature, in the dark. After two washing steps with PBS to remove the free WGA-FITC reagent, yeast cells were resuspended with PBS to a concentration of 0.5×10^7 cell/ml for FACS analysis.

Propidium iodide (PI) staining

- PI (Sigma) was freshly dissolved in PBS buffer to a final concentration of 1mg/ml as stock solution. For staining, yeast cells were suspended in PBS pH 7.2 to approximately 10^7 cell/ml and then, 3 μ l of PI stock solution was added into 1 ml yeast cell suspension. The sample was run within 5-10 minutes on a flow cytometer
25 (Becton Dickinson), which is capable of measuring red fluorescence (with a band pass filter >650). No washing steps were included.

Set-up of Becton Dickinson FACScan

- Analysis of FITC, PE and PI labelling of the cell population was accomplished at an
30 excitation wavelength of 488 nm, using a 15 mWatt argon ion laser. FITC emission was measured as a green signal (530 nm peak fluorescence) by the FL1 detector, PE was measured as an orange signal (575 nm peak fluorescence) by the FL2 detector, and PI was measured as a red signal (670 nm peak fluorescence) by the FL3 detector. The FACScan flow cytometer (Becton Dickinson) was operated

according to the standard protocol of the supplier. For multi-colour staining, electronic compensation was used among the fluorescence channels to remove residual spectral overlap. A minimum of 10,000 events was collected on each sample. Analysis of the multivariate data was performed with CELLQuest software
5 (Becton Dickinson Immunocytometry System).

Transformation and aging assay in nematode

The expression vector of human ferritin fragment (FTL) for *C. elegans* was derived from L4759 by replacing the GFP with FTL fragment.

10 Wild-type *C. elegans* strain (N2) was used as host for FTL expression. The animals were cultured and handled as described (Brenner, 1974). The transient overexpression of human FTL was carried out according to Jin (1999) using an Eppendorf FemtoJet-TransferMan NK injection system (Eppendorf, Leuven, Belgium). 25-30 worms were injected with plasmid carrying the human FTL gene or
15 control plasmid. Plasmid pRF4, which carries the dominant rol-6(su1006) allele was coinjected to mark transformed progeny. After a one-hour recovery period in M9 buffer, injected animals were allowed to lay eggs for approx. 40 hours on plates containing nematode growth medium (NGM) and a lawn of *E. coli* bacteria (OP50) as food. Transformed eggs were predominantly laid during the last 20 hours
20 resulting in a fairly synchronous experimental cohort. Subsequently, the injected animals were removed and progeny (F1) was allowed to grow at 24°C. Fourth stage larvae or young adults showing the Roller phenotype were transferred onto separate plates (NGM + OP50) containing 300 µM 5-fluoro-2'-deoxyuridine (FUDR, Sigma) to prevent progeny (F2) production. Live/dead scoring was carried out daily. Lifespan
25 is defined as the day when the first transformed larvae hatched until their death.

Construction of a full ferritin clone

A ferritin PCR fragment (end to stop codon) was generated from the hepatoma cDNA library by using specific primers (5'-ctacgagcgtctcctgaagatgc3' and
30 5'-cgcggatccaagtcgctgggctcagaaggctc-3'). This fragment was cloned directly into the TOPO vector (Invitrogen, The Netherlands) and then digested with NotI, generating a NotI fragment. Subsequently, the NotI fragment was inserted in the NotI site of ferritin light fragment clone (pGAL10-FL), resulting a 750 bp full ferritin clone in pSCGal-SN-10.

35

Example 1: Magnetic based sorting of yeast M-cells

To use yeast as an aging model, the first step needed is the development of a system, which allows the isolation of a relatively pure population of old yeast cells. The method for distinguishing and separation of *S. cerevisiae* cells between
5 generations is based on the fact that daughter cells have a wall that is newly formed and do not have any detectable wall remnants of the mother cells. Cells from an overnight culture of *S. cerevisiae* strain INVSc1 in minimal SD medium were covalently coated with biotin and designated as mother cells (M-cell). The M-cells were inoculated into fresh medium, and allowed to grow for 5-6 generations as
10 determined by the cell density that is measured by a UV-visible spectrophotometer (Shimadzu). After loading with anti-biotin beads, M-cells were sorted out using a magnetic sorter or MACS (Materials and Methods).

The purity of the collected M-cells was determined by staining with streptavidin-PE, which specifically binds to biotin coated on the cell wall of M-cells, followed by flow
15 cytometric analysis. Due to the reaction of biotin with streptavidin-PE, high density staining of biotinylated M-cells was shown. As show in Figure 2A, there was clear separation between stained M-cells and unstained daughter cells (D-cell) populations. Gate and marker were positioned to exclude D-cells from the M-cell population. In the layout of FSC versus SSC, as the matter of fact, the gated M-cells
20 mainly appeared at high FSC/SSC values representing a large cell size population (Fig. 2C) compared to a small cell population of D-cells which mainly located at lower FSC/SSC values (Fig. 2D). Statistic analysis showed that the purity of the isolated M-cells reached more than 85%. Figure 2B shows a PE staining performed on a depleted D-cell population, which hardly shows any positive signal.

25

Example 2: WGA based staining for analysis of yeast life span

Wheat germ agglutinin (WGA, *Triticum vulgare*) is the first lectin of which the amino acid sequence was completely determined (Wright, 1984). WGA is a mixture of several isolectins (Rice and Etzler, 1975). Sharing similar carbohydrate binding
30 properties with other lectins, WGA reacts strongly with the chitobiose core of asparagines linked oligosaccharides, especially with the Man β (1,4)GlcNAc β (1,4)GlcNAc trisaccharide (Yamamoto et al., 1981).

One of the most striking features of the cell surface during aging *S. cerevisiae* is the accumulation of chitin-containing bud scars. To verify whether WGA can be used for

specific labelling of chitin in yeast bud scars, the yeast strain INVSc-1(pEX2) was incubated with the FITC-conjugated WGA. The enriched, magnetically sorted M-cells were subjected to WGA reaction.

Under a fluorescence microscope we found that the major part of the fluorescent signal for WGA-FITC staining was co-localizing with the bud scar rings (Fig. 3). Moreover, the number of stained bud scars (6 bud scars) was consistent with the expected age of the M-cells as estimated by cell density measurement of the culture (5-6 generation). This observation demonstrated that, under the conditions used, WGA is specifically binding to the chitin of bud scars and hardly gives any fluorescence, caused by binding to compounds in the normal cell wall. Therefore, the possibility was examined to use WGA as a tool to stain bud scar for analysis of yeast life span. The isolated M-cells and depleted D-cells (as seen in Fig. 2C and 2D) were simultaneously stained with streptavidin-PE and WGA-FITC. As shown in Figure 2E-2F, D-cells that were negative for streptavidin-PE staining showed low FITC signal (Fig. 2F), whereas M-cells, which were positive in streptavidin-PE staining, showed a much stronger FITC staining (Fig. 2E). Under the fluorescent microscope, we observed that most M-cells contained 5-6 bud scar rings, which were strongly labelled by WGA-FITC, while most D-cells had only 1-2 bud scar rings. This observation indicated that there was a good linear correlation between the number of bud scars and the intensity of fluorescence. Therefore, it was assumed that WGA could be used as a tool for bud scar-specific staining in budding yeast cells.

Example 3: Application of using WGA to screen a human cDNA library

It has been reported that overexpression of certain human genes in yeast might have an influence in the frequency distribution of the yeast population (Gershon and Gershon, 2000). This overexpression of a single gene, which modulates the longevity in a single-cell system, has opened up the field of aging study to the power of yeast genetics. To screen human genes that might be involved in aging processes, a cDNA library from hepatoma cells was constructed and transferred into the yeast strain INVSc-1(pEX2) (See Materials and Methods). The transformed yeast population was first labelled with biotin and then cultured in a Bioreactor (AppliTek), for about 14 generations, as deduced from the cell density. According to the method described above, the initial biotinylated M-cells were isolated by

magnetic beads described herein and then labelled with WGA-FITC. By flow cytometric analysis (Fig. 4A), the M-cell population had a high density of WGA-FITC staining (gate M-cell), whereas D-cells showed a lower fluorescent staining (gate D-cell). As shown in Figure 4, older M-cells, gated as Old-M population, which were supposed to have a longer life span, were marked on high FITC intensity combined with high FSC, and then were flow sorted by FACS. From 9 colonies, the gene, overexpressed in the yeast cell was sequenced, and the results are summarized in Table 1. The growth rate was tested by measuring the doubling time of each strain in the liquid medium. The result showed that the growth rate of all 9 clones as well as the parent line were similar.

One of the colonies contained a gene fragment encoding ferritin light (FL) chain (M1147.1; Af119897.1). To verify whether the overexpression of this gene could influence the life-span of the yeast cell or not, an analysis of cell death using PI staining was performed in this ferritin L chain clone (CI2-FL) using its parent line of INVSc-1(pEX2) as a control. Ten million M-cells for each cell line were isolated. As shown in Figure 5, on the FSC versus PE (FL2) dot plot, a gate R2 was set around the PE-positive cells that represents the M-cell population while a gate R3 was set around the D-cell population. At the same time, on the FSC versus PI (FL3) dot plot, a gate R1 was set around PI-positive cells that cover the dead cells. As seen in Figure 5, cell death in culture occurred mainly in the M-cell population, but was barely detected in the D-cell population. Statistical analysis for dead cells (PI-positive) showed a higher frequency in control cells (33% death) compared to that in CI2-FL cells (16.3% death). This result indicates that over expression of human ferritin L chain in yeast cells prevents early cell death.

Example 4: additional screening experiments

To confirm the usefulness of the method, additional screening experiments were set up, using the same outline as described above both using the pEX2 library and the pSCGAL10-SN library. The results of the additional screening experiments are listed in Table 2, and identified by their genbank accession number. Several results of the first screening have been confirmed, illustrating the usefulness and the reliability of the method.

Example 5: protective effect of the ferritin fragment on hydrogen peroxide treatment

One of the colonies contained a gene fragment encoding ferritin light (FTL) chain (M1147.1; Af119897.1) cloned in pSCGAL10-SN. The plasmid was indicated as pGAL10-FL. Ferritin is ubiquitously distributed in the animal kingdom. It is composed of two subunits, the heavy chain (H) and the light chain (L). Ferritin plays a major role in the regulation of intracellular iron storage and homeostasis. One of the functions is to limit iron availability for participation in reactions that produce free oxygen radicals, which have the potential to damage lipids, proteins and DNA. Indeed, several reports have implicated that ferritin is involved in the protection against oxidative stress, such as stress induced by hydrogen peroxide. However, there is not such ferritin-like protein present in yeast, and anti-oxidative activity of ferritin fragments was never demonstrated. To test whether the human ferritin fragment plays a role as an antioxidant in yeast, we examined the partial-ferritin L clone (Cl2-ferritin), which was isolated by the method according to the invention, against H₂O₂ stress.

The condition for treatment of the cells was essentially the same as described by Jamieson et al. (1994). Exponential phase cultures of strain BY4741 that contained the empty vector pSCGAL10-SN (Control) and the ferritin expression vector (FTL – indicated as Cl2-ferritin) respectively, were grown aerobically in S-galactose medium at 30° C. The cell cultures were then challenged to a lethal concentration of H₂O₂ (1mM). Cell survival was monitored by taking samples at 0, 30 and 60 min, diluting the samples in the same medium and plating aliquots on YPD plates.

The experiment showed that, compared with control line, ferritin cells are significantly more resistant to treatment with 1mM H₂O₂ (Fig. 6).

Example 6: Transgenic nematode overexpressing the Ferritin Light chain

Although on the cellular level, there might be some conserved mechanism of aging processes throughout evolution (Martin et al., 1996), it is easy to imagine that in different species some underlying distinctive ways of intercellular regulation also contribute to reach their fate (Guarente 2001). In this sense, results from other organisms may provide a closer vision on the postulated function of human *FTL* gene involved in aging. Therefore, we tested whether FTL might affect lifespan in *C. elegans*, a multicellular organism, too. Indeed, as shown in Figure 5, animals

carrying human *FTL* genes appeared an average life of 13.5 days, which is 8% longer than the control line and statistically significant ($p=0.006$, two-way ANOVA). Many reports in *C.elegans*, *Drosophila* and mice are consistent with the hypothesis that oxidative damage accelerates aging, and that increased resistance to oxidative damage can extend lifespan (Finkel and Holbrook, 2000). The consistency that the expression/overexpression of human *FTL* gene was in favour in extending the lifespan in mono-cellular yeast and multi-cellular nematode supports the postulation that ferritin extends lifespan in cells, probably by protecting cells from oxidative stress, in a wide range of species.

A frequently practiced strategy in searching gene responsible for aging is by selecting survivals after exposure cells to stresses. Then a question constantly existing is that the genes picked up might be in response to the stress treatment rather than involved in aging, because of the complicity of the process. The screening method described here, however, provides an alternative that allows direct hunting of genes with potential anti-aging functions from various libraries or library combinations of eukaryotics. Yeast lines are selected in a more native condition, and also with advantages of high throughput, high efficiency, and short time consuming. Obviously, it has a great potential in application in rational drug design and therapies development in the field of age-related diseases preventing / treatments.

Example 7: elaboration of the mixed culture experiments

Based on the fact that a parental yeast strain and its direct derivative have a similar cell cycle rate, a mixed culture method has been developed to verify the long-living character of a transformed yeast strain when these strains are grown together in the same culture.

Two (or possibly more than two) yeast strains with a similar growth rate are initially mixed in the same culture in an equal ration (50% each in the case of two strains).

The strains can be distinguished from each other by the use of a selective marker.

The initial inoculated cells, called mother cells (M-cell), are labelled with biotin, and are grown together in the same culture during their entire life span. Mother cells at different generation points are sampled and collected by a magnetic system (MACS), similar to the method described in example 1. The ratio of living M-cells from the two strains is determined by the use of the selective marker. If the two strains have the

similar lifespan, the ratio of two viable strains will stay the same at different generation time points; otherwise, the ratio will change. This method is essentially based on the screening method, whereby the identification of the long living cells is not carried out by WGA staining, but by direct count of the number of living mother cells of the transformed strain(s), compared to the number of living mother cells of the parental strain.

FOB1 is required for the replication fork block. A *FOB1* mutation results in a decreased rDNA recombination rate and an increase in yeast life-span of 70%. The growth rate of the $\Delta fob1$ mutant strain, as measured, is similar to its parental strain.

Therefore, the long-living $\Delta fob1$ strain with its parental strain BY4742 was used to develop the mixed-growth system.

The initial mother cells were prepared as follows: a first pre-culture was made by inoculating BY4742 and $\Delta fob1$ cells (from freshly grown on a SD plate) in 5 ml of SD medium, respectively. The culture was incubated at 30 °C on a shaker at 250-300 rpm overnight. A second pre-culture was made by inoculating the first pre-culture into 5 ml of SD medium at a cell density of $OD_{600} = 0.001 \sim 0.005$. These cells were incubated until the culture reached a cell density of $OD_{600} = 0.5 \sim 0.7$. Cells were collected by centrifugation of the culture at 4 °C for 5 min at 3000 rpm. The cell pellet was washed twice with pre-cooled PBS (pH 8) and resuspended in PBS at a cell density of $OD_{600} = 5$ (approximately 5×10^7 cells/ml). The biotinylation of cells was performed in an eppendorf tube, in 1 ml reaction volume consisting of 0.5 ml of above-mentioned cells (2.5×10^7 cells) and 0.5 ml of 1 mg/ml biotin (Sulfo-NHS-LC-Biotin). The mixture was incubated for 30 min at room temperature with a gentle shaking. The biotinylated cells were centrifuged for 5 min at 13000 rpm and washed twice with 1 ml of cold PBS to get rid of free biotin. These cells were used as initial mother cells (M-cell).

A 100ml mixed-growth culture of BY4742 and $\Delta fob1$ was set up by inoculating 1×10^7 biotinylated M-cells from each strain (mother cells) at ratio of 1:1 in a SD medium. The mixed-growth culture was incubated at 30 °C on a shaker at 250-300 rpm. The culture density was not allowed to exceed $OD_{600} > 1$.

After growing several generations (up to 7-generation in our condition), the M-cells were labelled with anti-biotin microbeads and isolated using the magnetic system (MACS). The purity of M-cells was determined by FACS (fluorescence-activated cell sorter) after staining M-cells with streptavidin-conjugated with PE. Using these

conditions, more than 90% M-cells could be obtained. After the final magnetic sorting, the ratio of viable M-cells was measured.

Mixed M-cells samples were plated at about 500 cells per plate on YPD and YPD/geneticin plates to determine the ratio of mother cells of the two strains at different generation points. Plates were incubated for three days at 30 °C. The ratio of BY4742 and $\Delta fob1$ mother cells was monitored by counting the colonies on the two kind of plates. The total viable number of M-cells could be determined on the YPD plate, while the number of viable $\Delta fob1$ M-cells could be derived from YPD/geneticin plate.

As shown in Figure 8, the mixed M-cell group had similar amounts of the two strains at G0, while at G20 M-cells from $\Delta fob1$ were dominant (96%) among the cells sorted and collected with the magnetic sorting system. This result confirms that the mixed-growth method could indeed be used to distinguish the longer living yeast strain from its control.

Example 8: Confirmation of aging phenotype of ferritin strain by mixed-growth system

A kinetic analysis for growth rate of the ferritin yeast (FTL) and its parental strain INVSc-1 (with a geneticin-selectable marker) revealed a similar rate. About an equal amount of two strains was mixed, as described above, but using S-glycerol medium to obtain induction of the ferritin expression. This mixed culture was subjected to a mixed-growth experiment for determining their life span differences. After examination of the longevity of a mixed-growth of these two cell types by mixed-growth system and subsequent plating, we found that the ferritin line was predominant in the viable M-cell group after a growth of 10 generations (Figure 9). Growth of a mixture of these two lines in SD medium, in which the expression of ferritin not induced, revealed a constant viable FTL/INVSc-1 ratio. This indicates that the extended longevity of the FTL strain, compared to the age-matched INVSc-1 strain, is caused by the expression of human FTL.

Example 9: independent confirmation of effect on life span by ferritin

Iron is an essential nutrient for virtually every organism because is required as an essential cofactor for many proteins. However, excess iron can generate via the Fenton reaction highly toxic-free radicals generating oxidative damage to the cell.

Thus, cellular iron concentration must be tightly controlled. To exam whether expression of human ferritin in yeast could protect cell death upon excess iron, the lifespan analysis of ferritin strains was carried out by micromanipulaor as described previously (Kennedy et al., 1994) with the following slight modifications. Cells were

5 pregrown on non-inducing SD medium (2% glucose), shifted to inducing S-raffinose (2% raffinose) medium with 500 μ M ferric ammonium citrate (FAC) and 80 μ M ferrichrome (Sigma), and grown for at least two generations. Cells were taken from this logarithmically growing liquid culture and transferred at low density on S-

10 raffinose with 500 μ M FAC and 80 μ M ferrichrome plate (2% agar). The cells were then incubated at 30 °C overnight. Virgin daughters cells were isolated as buds from populations by micromanipulator and used as the starting mother cells for life span analysis. For each successive bud removed from these mother cells, they were counted one generation older. Cells were grown at 30°C during the day and at

15 cold room overnight. Each experiment consists of at least 60 cells. The statistical analysis of life span was carried by a Wilcoxin 's test. The life span of full ferritin and partial ferritin yeast strains were significantly extended by 10 to 15% compared to their parent strain BY4741 (Figure 10). This result confirms that human ferritin light chain prevents fast aging in presence of iron in yeast.

20 **Table 1:** results of the screening of 9 positive clones

Clone number	Insert length (approx.)	Identification (Based on homology)	SEQ ID N°
1	1.6 kb	Humanin	1
2	883 bp	APOA1	3
3	1 kb	Ribosomal protein P0	5
4	2.8 kb	glutamyl tRNA synthetase	7 ⁽¹⁾
5	2.4 kb	GRSF-1	8 ⁽¹⁾
7	700 bp	ALDH1	9
8	416 bp	ferritin light chain	11
9	1 kb	Ribosomal protein S2	13
12	500 bp	Histone H2A	15 ⁽¹⁾

(1) antisense

Table 2: Results of further screening experiments. The results are grouped in mitochondrial functions, ribosomal proteins, other genes with known function, unknown functions and chromosomal fragments. The results of the first screening are not repeated in this table; however, several genes, like the ferritin fragment, have been identified in more than one screening experiment. The sequences are identified by their genbank accession number. The length of the isolated fragment may differ from the genbank sequence, and is normally shorter. Where relevant, the fragment is indicated, using the nucleotides numbers of the genbank sequence.

Mitochondrion

clone name	Function	accession number	orientation
1E3/6D8	ATP synthase 6 mRNA,	AF368271	sense
2C10	mitochondrial ATP synthase subunit 9, P3 gene copy, mRNA, nuclear gene encoding mitochondrial protein	U09813	sense
5D9	ATP synthase, H ⁺ transporting, mitochondrial F1, complex, O subunit (oligomycin sensitivity conferring protein), (ATP5O)	NM_001697 89-745	sense
9B11	ADP/ATP translocase mRNA, 3' end	J03591	sense
4D7/7H1/12D3 /13E9	NADH dehydrogenase 1	BC009316 380-685; 10-684; 138-645; 10-490	sense
6C11	NADH dehydrogenase 1	BC009316	sense
7E11	NADH dehydrogenase subunit 5 (MTND5) mRNA, RNA 4, complete cds; mitochondrial gene for mitochondrial product	AF339086	sense
10G3	mitochondrion cytochrome b gene, partial cds	U09500	sense
12F1	cytochrome c oxidase subunit III gene, mitochondrial gene encoding mitochondrial protein, partial cds	AF004341	sense
2A7	ubiquinol-cytochrome c reductase core protein II	BC003136; 763-1331	sense
1B12	monocyte chemotactic protein-3 (MCP-3)	X72308	
1F9/12H12	Wnt-13 ; metochrondrial DNA	Z71621; 1-348; 12-372	sense
7C1	12S ribosomal RNA gene, partial sequence; and tRNA-Val gene, complete sequence; mitochondrial genes for mitochondrial products	AY012136	sense

11B7	MRPS16 mRNA for mitochondrial ribosomal protein S16	AB049948.	sense
10F3/ 14H4/14H5 /7B10	clone IMAGE:5581122, mRNA; haplotype N1b mitochondrion	<u>BC035832.1</u> <u>AF381999</u> ; <u>228-726; 330-953 ; 134-</u> <u>1028 2059-2658</u>	sense

Ribosome

1S_3	ribosomal protein P0	BC005863	sense
3A5	ribosomal protein, large, P1	NM_001003.2	sense
12E12	ribosomal protein L12 (RPL12)	NM_000976	sense
6F8	ribosomal protein L14	BC029036	sense
1D12	ribosomal protein L31 (RPL31)	NM_000993	sense
1S_9	ribosomal protein S2	NM_002952	sense
6D6	ribosomal S3 (RPS3);	NM_001005.2	sense
3D1/4G10	ribosomal protein S3A	<u>BC030161</u>	sense
	v-fos transformation effector protein (Fte-1)	M84711	sense
3B9	ribosomal protein S4, X-linked (RPS4X)	<u>NM_001007</u>	sense
	scar protein	M22146	sense
4H2	ribosomal protein S4, Y-linked (RPS4Y)	<u>NM_001008</u>	sense
10E8	ribosome protein S5	BC018151	sense
14G6	ribosomal protein S6 (RPS6)	NM_001010.2	sense
4C5	ribosomal protein S10,	<u>BC005012</u>	sense
4B5/2A3	ribosomal protein S11	BC016378	sense
	Mus musculus RAD21 homolog (S. pombe) (Rad21)	NM_009009	sense
11E4	ribosome protein S16	nm_001020	sense
2E6	ribosomal protein S17 mRNA	<u>M13932</u>	sense
1D1	ribosomal protein S25	<u>BC004986</u>	sense
1C11	Wilm's tumor-related protein (QM) mRNA; RPL10	M64241	sense

Other genes from the 4th screen (pEX2 library)**Unknown functions**

2H4	likely ortholog of mouse gene rich cluster, C8	<u>NM_031299.2</u> 346- <u>end</u>	sense
3C2	clone FLC0593	<u>AF113701</u>	sense
4C11	similar to putative, clone MGC:33177 IMAGE:4823662	BC028387; 1905-end	sense
4D10	full length insert cDNA clone ZE03C06	<u>AF086514</u>	sense
4E9	hypothetical protein dJ465N24.2.1 (DJ465N24.2.1)	NM_020317; 874-1431	sense
6F6	Similar to RIKEN cDNA 1110012M11 gene	BC007883	sense

6H8	cDNA FLJ31039 fis, clone HSYRA2000221	AK055601; 1869-end	sense
7F6	cDNA FLJ13305 fis	AK023367	sense
8C10	hypothetical protein FLJ23018 (FLJ23018)	NM_024810	sense
9F4	Similar to hypothetical protein FLJ10751	<u>BC024001; 3-end</u>	sense
9G10	hypothetical protein BC013073 (LOC92703)	NM_138391	sense
10G6	similar to C50F4.16.p (LOC256281)	XM_170755	antisense
12E6	hypothetical protein MGC955	NM_024097.1	sense
14D4	clone IMAGE; 4778940 mRNA	<u>BC031919.1; 3-end</u>	sense
5S-15/114	cDNA DKFZp434O159	AL133593	sense
5S-21/57	hypothetical protein FLJ10081	NM_017991	sense
7F11	cDNA FLJ38528 fis	AK095847	sense
11H3	cDNA FLJ14279 fis	AK024341; 362-end	sense
5C4	Similar to KIAA0674 protein	BC026048	sense
2E2	cDNA FLJ14385 fis, clone HEMBA1002212, weakly similar to TYROSINE-PROTEIN KINASE 2	AK027291; 3-end	sense
7G6	KIAA0776 protein (KIAA0776)	NM_015323; 3-end	sense

Chromosome DNA seq.

10D4	DNA sequence from clone RP1-64K7 on chromosome 20q11.21-11.23 Contains the EIF2S2 gene for eukaryotic translation initiation factor 2 subunit 2 (beta, 38kD), a putative novel gene, the gene for heterogenous nuclear ribonucleoprotein RALY or autoantigen P542, an RPS2 (RPS4) (40S ribosomal protein S2) pseudogene, ESTs, STS, GSSs and two CpG islands	<u>AL031668; 66383-66970</u>	sense
2A6	PAC clone RP3-414A15 from 14q24.3	<u>AC005225; 93459-93782</u>	sense
2D3	DNA sequence from clone RP11-357H24 on chromosome 10	AL451084; 42698-42510; with polyA	antisense
2F8	chromosome 17, clone hRPC.1110_E_20	<u>AC004231; 42223-429716; with polyA</u>	sense
3F11	BAC clone CTD-2314H8	<u>AC079338; 21007-21487</u>	sense
4F7	chromosome 1 clone RP11-109I2	AC091609; 155982-156333	sense
11B8	DNA sequence from clone RP11-735A5 on chromosome 1	AL603888	antisense
12A9	chromosome 18, clone RP11-13N13	AC106037.9	sense

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25

Claims

1. Method to screen genes involved in aging and/or in AAD's and/or in oxidative stress, comprising a) mutation or transformation of a yeast cell b) cultivation of said cell c) enrichment of the population for mother cells d) labelling said mother cells with a WGA based label and e) isolation of the highly labelled cells.
2. A method according to claim 1, whereby said WGA based label is FITC-conjugated WGA.
3. A method according to claim 1 or 2, whereby said isolation is a FACS based sorting.
4. A method according to any of the preceding claims, whereby said enrichment is a magnetic-based sorting.
5. A method according to any of the preceding claims, whereby said transformation is carried out with a yeast expression library.
6. A method according to claim 5, whereby said yeast expression library is expressing mammalian DNA or plant DNA.
7. A gene or functional gene fragment isolated with a method according to any of the claims 1-6.
8. A gene or functional gene fragment according to claim 7, comprising SEQ ID N°1, 3, 5, 7, 8, 9, 11, 13, 15, 16, 17, 19, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52 or 53.
9. The use of a gene or functional gene fragment, according to claim 7 or 8, to modulate aging and/or to protect against oxidative stress.
10. The use according to claim 9, whereby said gene comprises SEQ ID N° 11 or 16.
11. A polypeptide, encoded by a functional gene fragment according to claim 7.
12. The use of a polypeptide, encoded by a gene or functional gene fragment according to claim 7, to modulate aging and/or to protect against oxidative stress.
13. The use of a polypeptide, according to claim 11, whereby said gene or functional gene fragment comprises SEQ ID N° 2, 4, 6, 10, 14, 18 or 20.
14. The use of a polypeptide, according to claim 11, whereby said polypeptide comprises SEQ ID N° 12.
15. The use of a polypeptide, according to claim 11, whereby said polypeptide is encoded by SEQ ID N° 16.

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Fig. 1

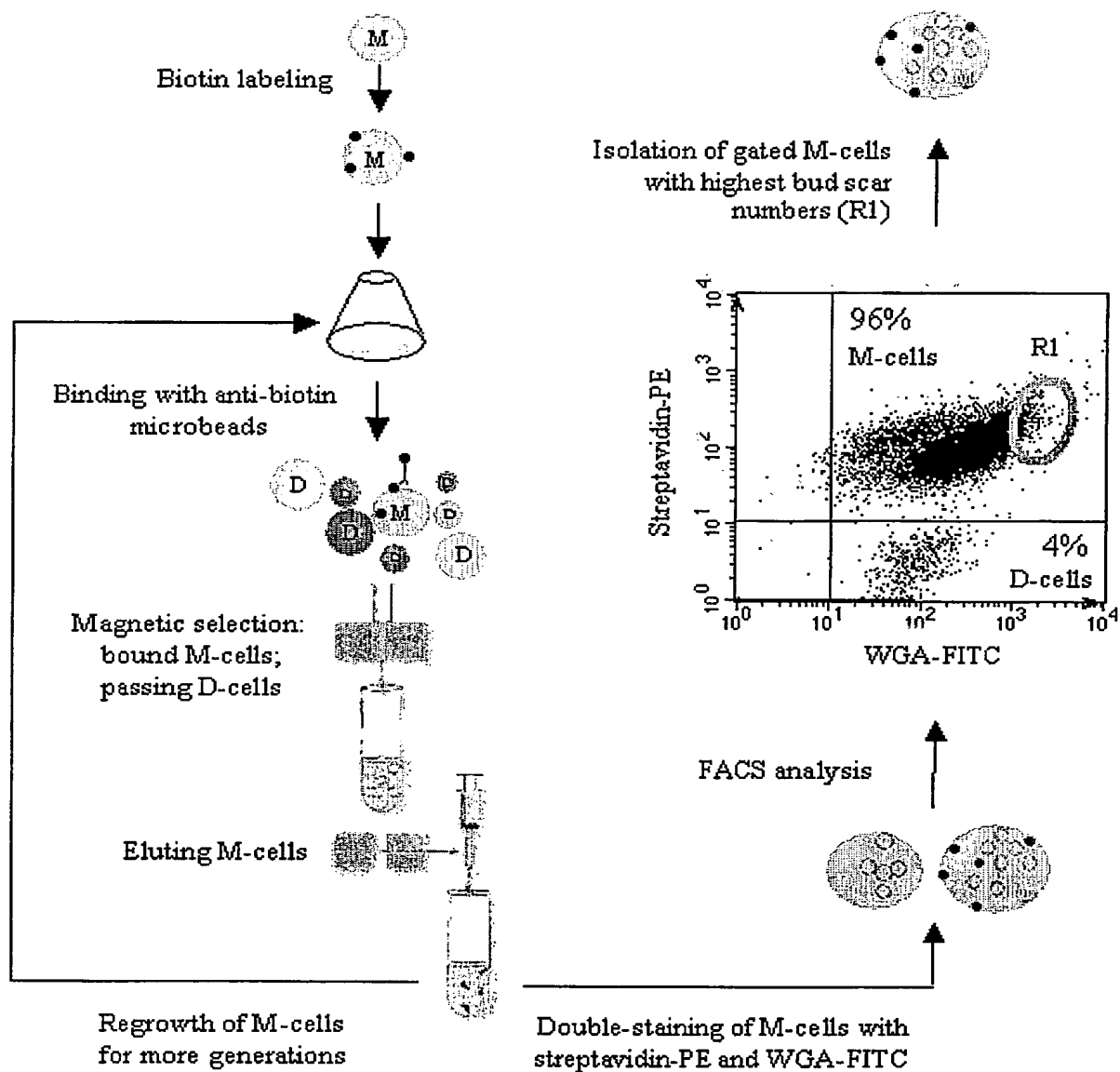
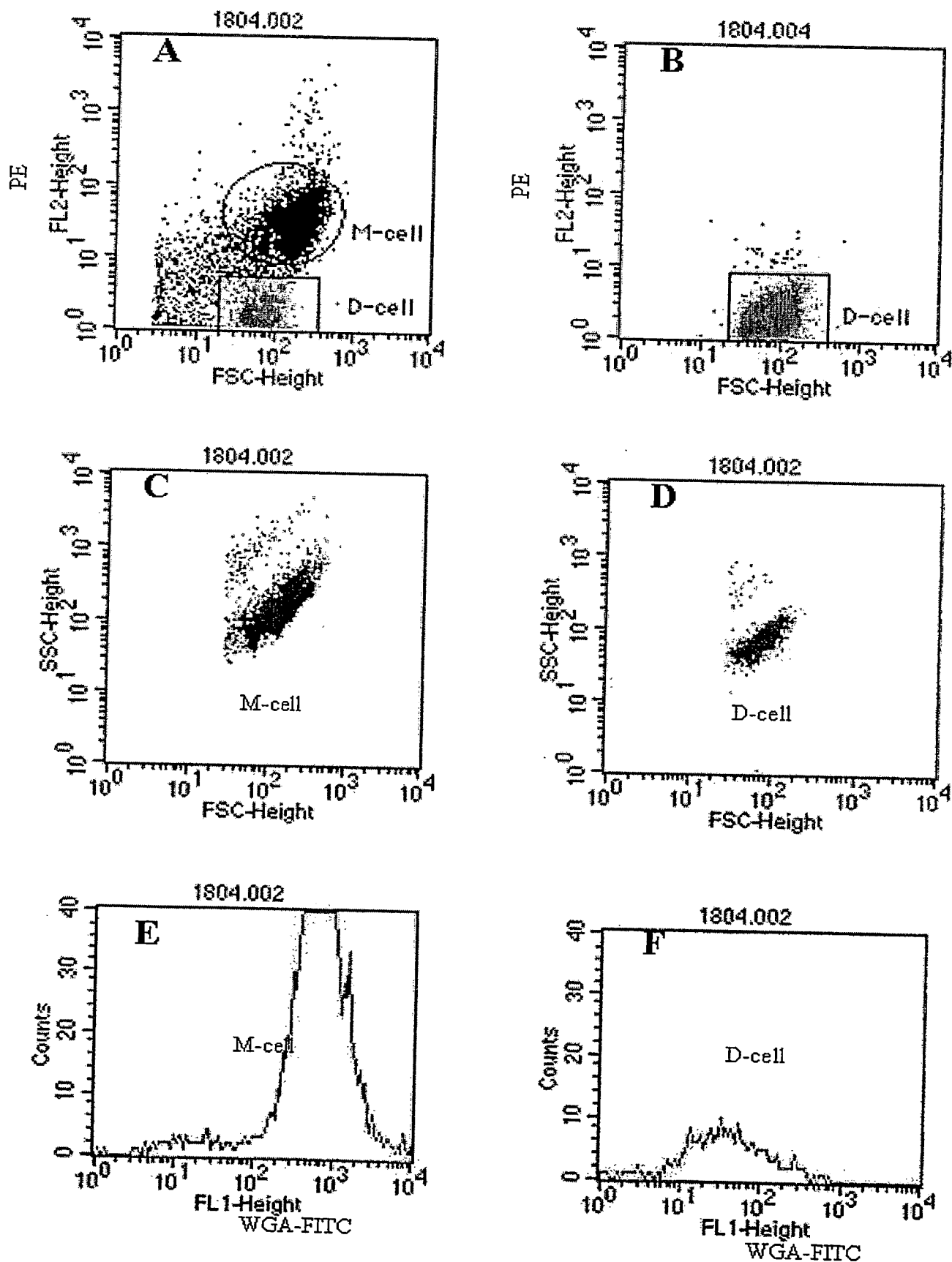


Fig. 2



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Fig. 3

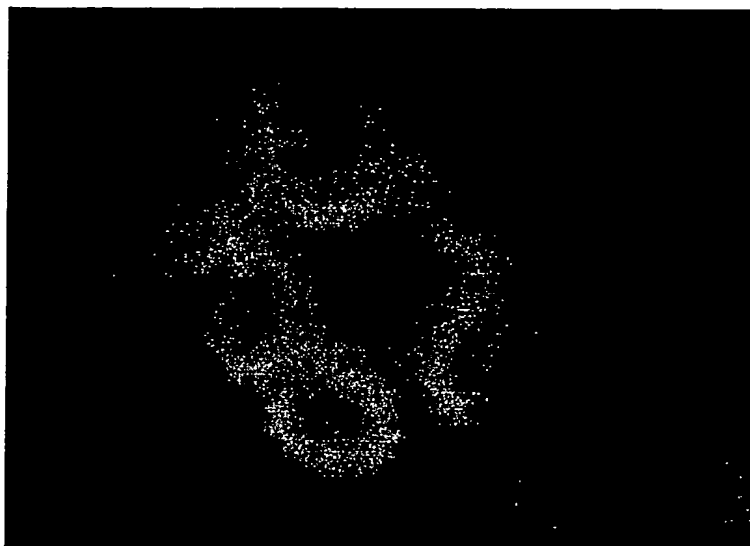
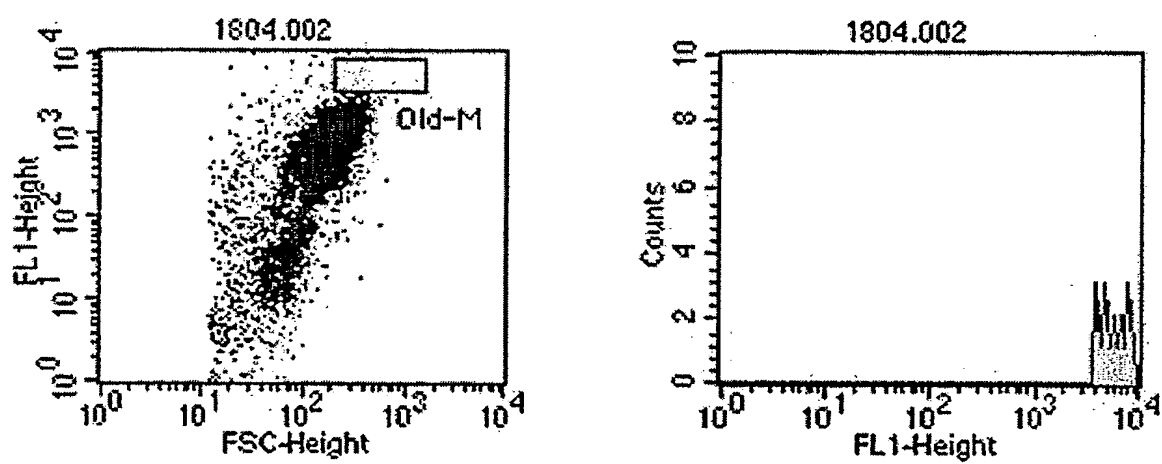
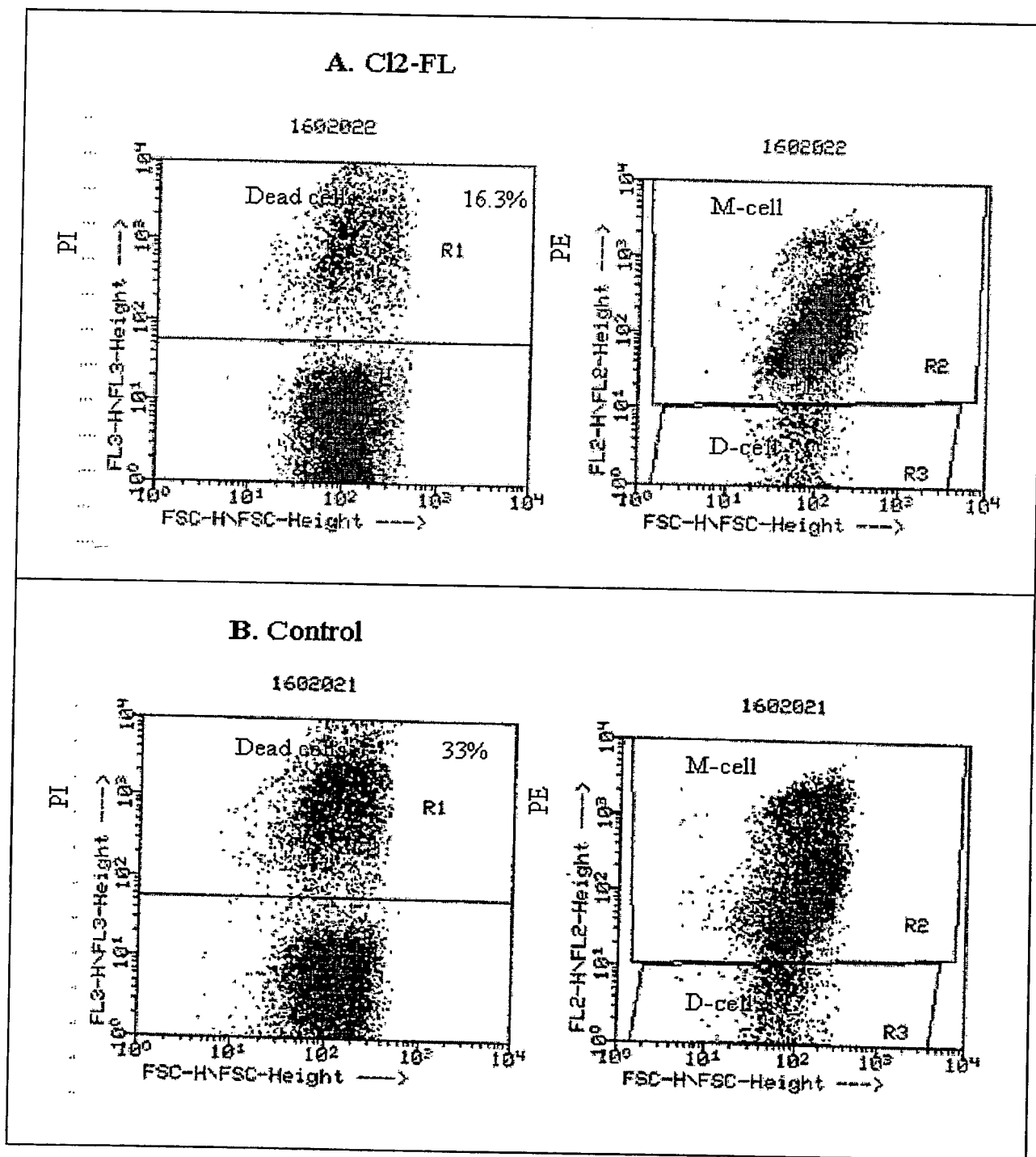


Fig. 4



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Fig. 5



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Fig. 6

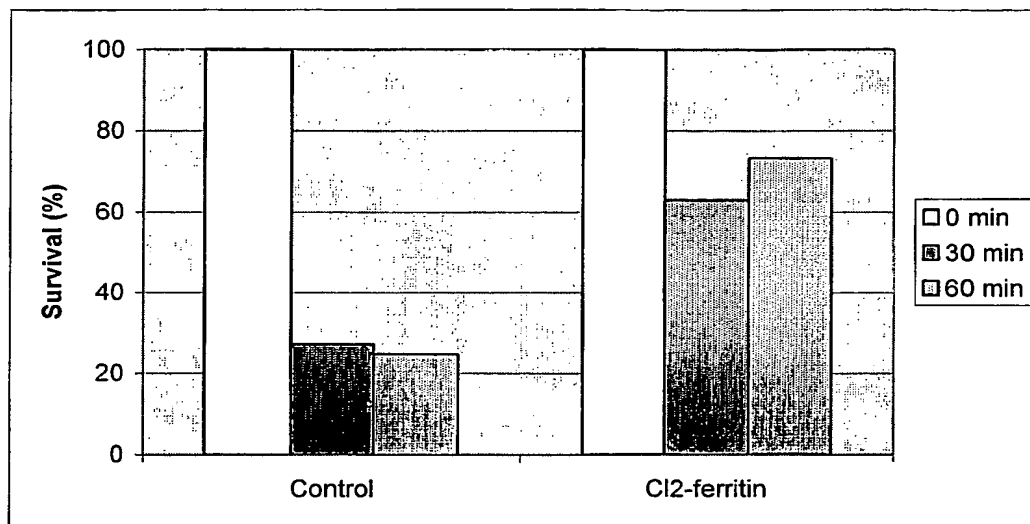
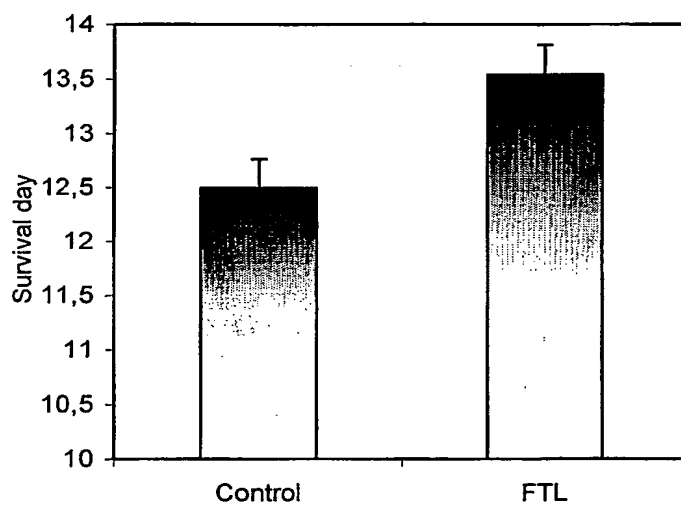


Fig. 7



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Fig. 8

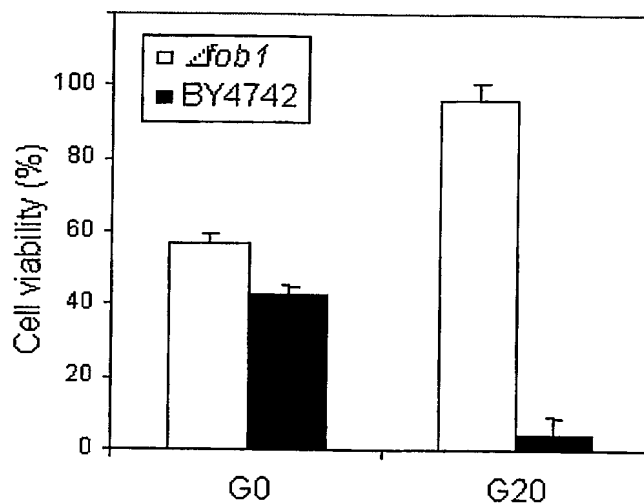
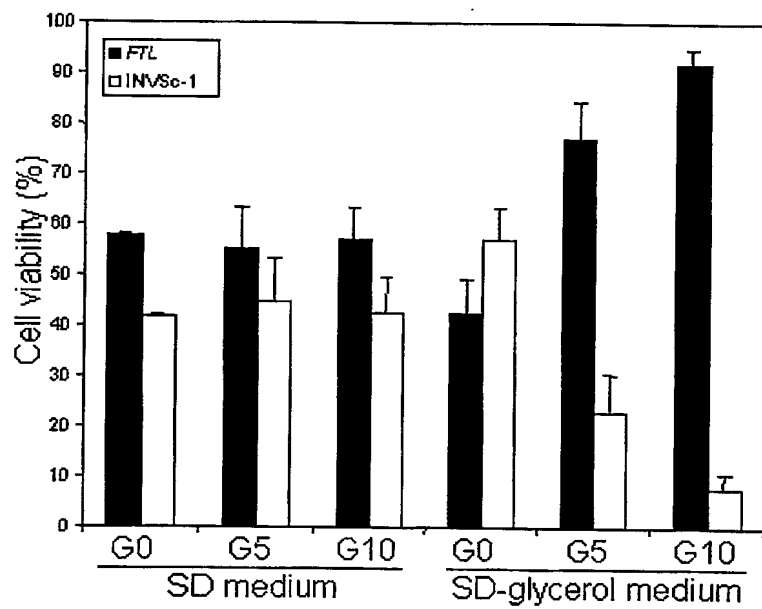
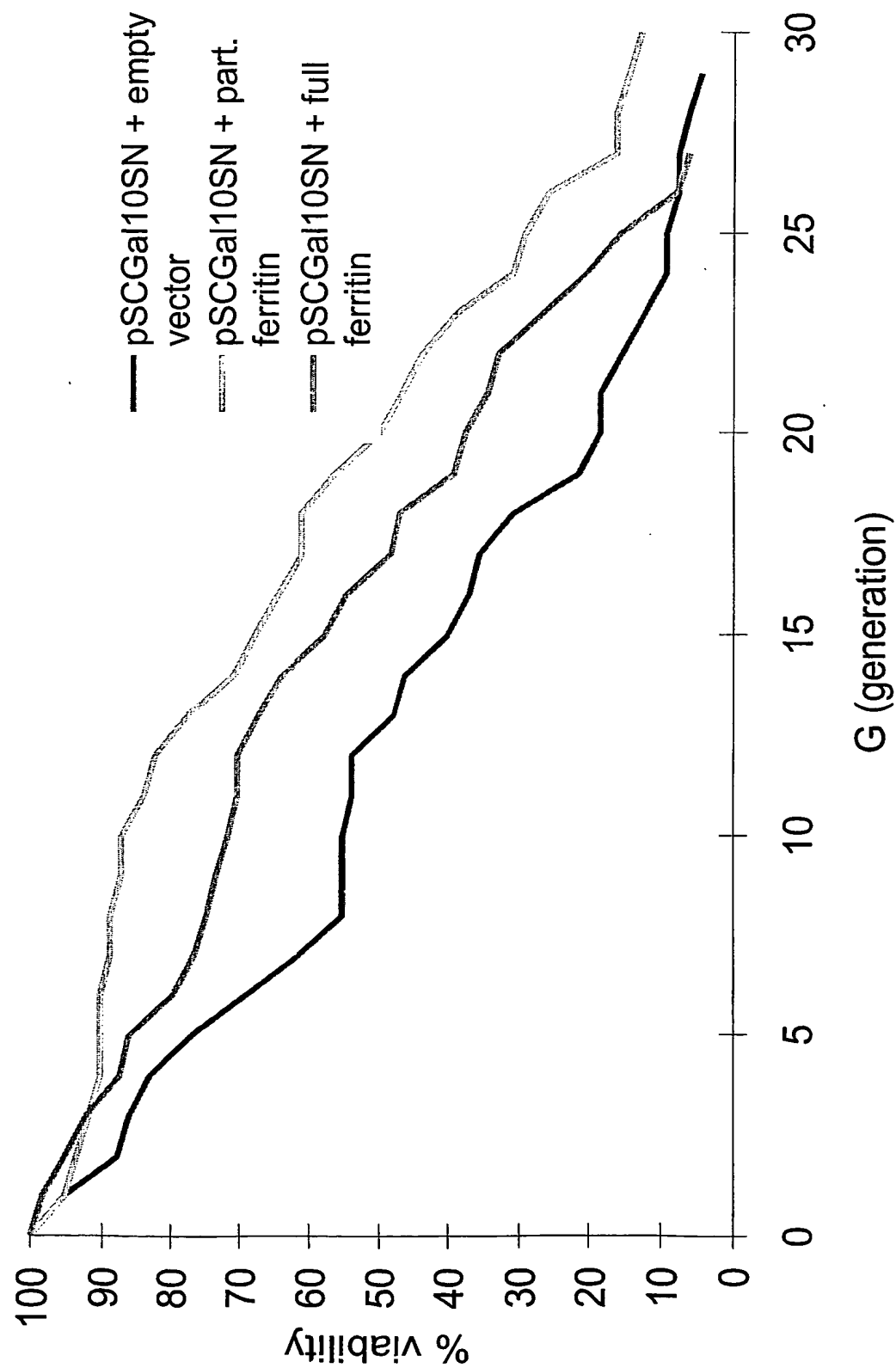


Fig. 9



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Fig. 10



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<210> 8

<211> 2233

<212> DNA

<213> Homo sapiens

<220>

<223> human G rich sequence factor antisense cDNA

<400> 8

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<220>

<221> CDS

<222> (2)..(55)

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1

5

10

15

tca taa agaaaataca agagtggaga gaagctcttc aatagctaag catctcctta 105
 Ser

cagtcactaa tatagtagat tttaaagaca aaatTTTTtct tttcttgatt tttttaaaca 165

taagctaaat catattagta ttaatactac ccatagaaaa cttgacatgt agcttcttct 225

gaaagaatta tttgccttct gaaatgtgac cccaagtcc tatcctaaat aaaaaaagac 285

aaattcggat gtatgatctc tctagctttg tcatagttat gtgattttcc tttgtagcta 345

cttttgcagg ataataattt tatagaaaag gaacagttgc atttagcttc tttcccttag 405

tgactcttga agtacttaac atacacgtta actgcagagt aaattgctct gttcccagta 465

gttataaagt cttggactg ttttgaaaag tttcctagga tgtcatgtct gottgtcaaa 525

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tgcatgat 593

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 <213> Homo sapiens
 <223> human ALDH1 cDNA fragment

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 Ser

<210> 11
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 <212> DNA
 <213> Homo sapiens

<220>
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<220>
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 <222> (2)..(418)

<400> 11

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    1             5             10             15

tcc ggg gac tct ctt cca gcc tcc gac cgc cct ccg att tcc tct ccg 97
  Ser Gly Asp Ser Leu Pro Ala Ser Asp Arg Pro Pro Ile Ser Ser Pro
           20             25             30

ctt gca acc tcc ggg acc atc ttc tcg gcc atc tcc tgc ttc tgg gac 145
  Leu Ala Thr Ser Gly Thr Ile Phe Ser Ala Ile Ser Cys Phe Trp Asp
           35             40             45

ctg cca gca ccg ttt ttg tgg tta gct cct tct tgc caa cca acc atg 193
  Leu Pro Ala Pro Phe Leu Trp Leu Ala Pro Ser Cys Gln Pro Thr Met
           50             55             60

agc tcc cag att cgt cag aat tat tcc acc gac gtg gag gca gcc gtc 241
  Ser Ser Gln Ile Arg Gln Asn Tyr Ser Thr Asp Val Glu Ala Ala Val
        65             70             75             80

aac agc ctg gtc aat ttg tac ctg cag gcc tcc tac acc tac ctc tct 289
  Asn Ser Leu Val Asn Leu Tyr Leu Gln Ala Ser Tyr Thr Tyr Leu Ser
           85             90             95

ctg ggc ttc tat ttc gac cgc gat gat gtg gct ctg gaa ggc gtg agc 337
  Leu Gly Phe Tyr Phe Asp Arg Asp Asp Val Ala Leu Glu Gly Val Ser
           100             105             110

cac ttc ttc cgc gaa ttg gcc gag gag aag cgc gag ggc tac gag cgt 385
  His Phe Phe Arg Glu Leu Ala Glu Glu Lys Arg Glu Gly Tyr Glu Arg
           115             120             125

ctc ctg aag atg caa aac cag cgt ggc ggc cgc 418
  Leu Leu Lys Met Gln Asn Gln Arg Gly Gly Arg
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<210> 12

<211> 139

<212> PRT

<213> Homo sapiens

<223> human ferritin cDNA fragment

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Ser Gly Asp Ser Leu Pro Ala Ser Asp Arg Pro Pro Ile Ser Ser Pro

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20 25 30
 Leu Ala Thr Ser Gly Thr Ile Phe Ser Ala Ile Ser Cys Phe Trp Asp
 35 40 45
 Leu Pro Ala Pro Phe Leu Trp Leu Ala Pro Ser Cys Gln Pro Thr Met
 50 55 60
 Ser Ser Gln Ile Arg Gln Asn Tyr Ser Thr Asp Val Glu Ala Ala Val
 65 70 75 80
 Asn Ser Leu Val Asn Leu Tyr Leu Gln Ala Ser Tyr Thr Tyr Leu Ser
 85 90 95
 Leu Gly Phe Tyr Phe Asp Arg Asp Asp Val Ala Leu Glu Gly Val Ser
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<211> 929

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<213> Homo sapiens

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<220>

<221> CDS

<222> (11)..(892)

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 Pro Gly Gly Pro Gly Met Gly Asn Arg Gly Gly Phe Arg Gly Gly Phe
 15 20 25
 ggc agt ggc atc cgg ggc cgg ggt cgc ggc cgt gga cgg ggc cgg ggc 145
 Gly Ser Gly Ile Arg Gly Arg Gly Arg Gly Arg Gly Arg Gly

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cga ggc cgc gga gct cgc gga ggc aag gcc gag gat aag gag tgg atg				193
Arg Gly Arg Gly Ala Arg Gly Gly Lys Ala Glu Asp Lys Glu Trp Met				
	50	55	60	
ccc gtc acc aag ttg ggc cgc ttg gtc aag gac atg aag atc aag tcc				241
Pro Val Thr Lys Leu Gly Arg Leu Val Lys Asp Met Lys Ile Lys Ser				
	65	70	75	
ctg gag gag atc tat ctc ttc tcc ctg ccc att aag gaa tca gag atc				289
Leu Glu Glu Ile Tyr Leu Phe Ser Leu Pro Ile Lys Glu Ser Glu Ile				
	80	85	90	
att gat ttc ttc ctg ggg gcc tct ctc aag gat gag gtt ttg aag att				337
Ile Asp Phe Phe Leu Gly Ala Ser Leu Lys Asp Glu Val Leu Lys Ile				
	95	100	105	
atg cca gtg cag aag cag acc cgt gcc ggc cag cgc acc agg ttc aag				385
Met Pro Val Gln Lys Gln Thr Arg Ala Gly Gln Arg Thr Arg Phe Lys				
	110	115	120	125
gca ttt gtt gct atc ggg gac tac aat ggc cac gtc ggt ctg ggt gtt				433
Ala Phe Val Ala Ile Gly Asp Tyr Asn Gly His Val Gly Leu Gly Val				
	130	135	140	
aag tgc tcc aag gag gtg gcc acc gcc atc cgt ggg gcc atc atc ctg				481
Lys Cys Ser Lys Glu Val Ala Thr Ala Ile Arg Gly Ala Ile Ile Leu				
	145	150	155	
gcc aag ctc tcc atc gtc ccc gtg cgc aga ggc tac tgg ggg aac aag				529
Ala Lys Leu Ser Ile Val Pro Val Arg Arg Gly Tyr Trp Gly Asn Lys				
	160	165	170	
atc ggc aag ccc cac act gtc cct tgc aag gtg aca ggc cgc tgc ggc				577
Ile Gly Lys Pro His Thr Val Pro Cys Lys Val Thr Gly Arg Cys Gly				
	175	180	185	
tct gtg ctg gta cgc ctc atc cct gca ccc agg ggc act ggc atc gtc				625
Ser Val Leu Val Arg Leu Ile Pro Ala Pro Arg Gly Thr Gly Ile Val				
	190	195	200	205
tcc gca cct gtg cct aag aag ctg ctc atg atg gct ggt atc gat gac				673
Ser Ala Pro Val Pro Lys Lys Leu Leu Met Met Ala Gly Ile Asp Asp				
	210	215	220	
tgc tac acc tca gcc cgg ggc tgc act gcc acc ctg ggc aac ttc gcc				721
Cys Tyr Thr Ser Ala Arg Gly Cys Thr Ala Thr Leu Gly Asn Phe Ala				

225	230	235	
aag gcc acc ttt gat gcc att tct aag acc tac agc tac ctg acc ccc			769
Lys Ala Thr Phe Asp Ala Ile Ser Lys Thr Tyr Ser Tyr Leu Thr Pro			
240	245	250	
gac ctc tgg aag gag act gta ttc acc aag tct ccc tat cag gag ttc			817
Asp Leu Trp Lys Glu Thr Val Phe Thr Lys Ser Pro Tyr Gln Glu Phe			
255	260	265	
act gac cac ctc gtc aag acc cac acc aga gtc tcc gtg cag cgg act			865
Thr Asp His Leu Val Lys Thr His Thr Arg Val Ser Val Gln Arg Thr			
270	275	280	285
cag gct cca gct gtg gct aca aca tag ggtttttata caagaaaaat			912
Gln Ala Pro Ala Val Ala Thr Thr			
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<210> 14
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 <212> PRT
 <213> Homo sapiens
 <223> human ribosomal protein S2 cDNA

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 35 40 45
 Gly Ala Arg Gly Gly Lys Ala Glu Asp Lys Glu Trp Met Pro Val Thr
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 Lys Leu Gly Arg Leu Val Lys Asp Met Lys Ile Lys Ser Leu Glu Glu
 65 70 75 80
 Ile Tyr Leu Phe Ser Leu Pro Ile Lys Glu Ser Glu Ile Ile Asp Phe
 85 90 95
 Phe Leu Gly Ala Ser Leu Lys Asp Glu Val Leu Lys Ile Met Pro Val
 100 105 110
 Gln Lys Gln Thr Arg Ala Gly Gln Arg Thr Arg Phe Lys Ala Phe Val
 115 120 125
 Ala Ile Gly Asp Tyr Asn Gly His Val Gly Leu Gly Val Lys Cys Ser
 130 135 140
 Lys Glu Val Ala Thr Ala Ile Arg Gly Ala Ile Ile Leu Ala Lys Leu
 145 150 155 160

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Ser Ile Val Pro Val Arg Arg Gly Tyr Trp Gly Asn Lys Ile Gly Lys
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Pro His Thr Val Pro Cys Lys Val Thr Gly Arg Cys Gly Ser Val Leu
      180                      185                      190
Val Arg Leu Ile Pro Ala Pro Arg Gly Thr Gly Ile Val Ser Ala Pro
      195                      200                      205
Val Pro Lys Lys Leu Leu Met Met Ala Gly Ile Asp Asp Cys Tyr Thr
      210                      215                      220
Ser Ala Arg Gly Cys Thr Ala Thr Leu Gly Asn Phe Ala Lys Ala Thr
      225                      230                      235                      240
Phe Asp Ala Ile Ser Lys Thr Tyr Ser Tyr Leu Thr Pro Asp Leu Trp
      245                      250                      255
Lys Glu Thr Val Phe Thr Lys Ser Pro Tyr Gln Glu Phe Thr Asp His
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Leu Val Lys Thr His Thr Arg Val Ser Val Gln Arg Thr Gln Ala Pro
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Ala Val Ala Thr Thr
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<211> 525

<212> DNA

<213> Homo sapiens

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<223> human H2A histone antisense cDNA

<400> 15

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gccaacggag gctcagtcga gacagggatt aaccgacttg tgctggtatc taggtgcttg 180
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gagggagagc tgatgtgaaa ggccctgggtc ccggccgcag aaggggagcc agaccgaaat 480
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<210> 16

<211> 1381

<212> DNA

<213> Homo sapiens

<400> 16

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<220>

<221> CDS

<222> (75)..(581)

<400> 17

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ccaacctatt atat atg ccg ctc cta cga gga ctg ctg tgg ctc cag gtg 110
          Met Pro Leu Leu Arg Gly Leu Leu Trp Leu Gln Val
                1                      5                      10

ctg tgt gcg ggc cct ctc cat aca gag gct gtg gta ctt ctg gtt cct 158
Leu Cys Ala Gly Pro Leu His Thr Glu Ala Val Val Leu Leu Val Pro
        15                      20                      25

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tct gat gat ggg cgt gct ttt ctg ctg cgg agc cgg ctt ctt cat ccg 206
 Ser Asp Asp Gly Arg Ala Phe Leu Leu Arg Ser Arg Leu Leu His Pro
 30 35 40

gag gcg cat gta ccc ccc gcc gct gat cga gga gcc agc ctt caa tgt 254
 Glu Ala His Val Pro Pro Ala Ala Asp Arg Gly Ala Ser Leu Gln Cys
 45 50 55 60

gtc cta cac cag gca gcc ccc aaa tcc cgg ccc agg agc cca gca gcc 302
 Val Leu His Gln Ala Ala Pro Lys Ser Arg Pro Arg Ser Pro Ala Ala
 65 70 75

ggg gcc gcc cta tta cac cga ccc agg agg acc ggg gat gaa ccc tgt 350
 Gly Ala Ala Leu Leu His Arg Pro Arg Arg Thr Gly Asp Glu Pro Cys
 80 85 90

cgg gaa ttc cat ggc aat ggc ttt cca ggt ccc acc caa ctc acc cca 398
 Arg Glu Phe His Gly Asn Gly Phe Pro Gly Pro Thr Gln Leu Thr Pro
 95 100 105

ggg gag tgt ggc ctg ccc gcc ccc tcc agc cta ctg caa cac gcc tcc 446
 Gly Glu Cys Gly Leu Pro Ala Pro Ser Ser Leu Leu Gln His Ala Ser
 110 115 120

gcc ccc gta cga aca ggt agt gaa ggc caa gta gtg ggg tgc cca cgt 494
 Ala Pro Val Arg Thr Gly Ser Glu Gly Gln Val Val Gly Cys Pro Arg
 125 130 135 140

gca aga gga gag aca gga gag ggc ctt tcc ctg gcc ttt ctg tct tcg 542
 Ala Arg Gly Glu Thr Gly Glu Gly Leu Ser Leu Ala Phe Leu Ser Ser
 145 150 155

ttg atg ttc act tcc agg aac ggt ctc gtg ggc tgc taa gggcagttcc 591
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 160 165

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<211> 168

<212> PRT

<213> Homo sapiens

<223> clone 11A12 - hypothetical protein DKFZp564K0822

<400> 18

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Pro	Leu	His	Thr	Glu	Ala	Val	Val	Leu	Leu	Val	Pro	Ser	Asp	Asp	Gly	20	25	30	
Arg	Ala	Phe	Leu	Leu	Arg	Ser	Arg	Leu	Leu	His	Pro	Glu	Ala	His	Val	35	40	45	
Pro	Pro	Ala	Ala	Asp	Arg	Gly	Ala	Ser	Leu	Gln	Cys	Val	Leu	His	Gln	50	55	60	
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Leu	His	Arg	Pro	Arg	Arg	Thr	Gly	Asp	Glu	Pro	Cys	Arg	Glu	Phe	His	85	90	95	
Gly	Asn	Gly	Phe	Pro	Gly	Pro	Thr	Gln	Leu	Thr	Pro	Gly	Glu	Cys	Gly	100	105	110	
Leu	Pro	Ala	Pro	Ser	Ser	Leu	Leu	Gln	His	Ala	Ser	Ala	Pro	Val	Arg	115	120	125	
Thr	Gly	Ser	Glu	Gly	Gln	Val	Val	Gly	Cys	Pro	Arg	Ala	Arg	Gly	Glu	130	135	140	
Thr	Gly	Glu	Gly	Leu	Ser	Leu	Ala	Phe	Leu	Ser	Ser	Leu	Met	Phe	Thr	145	150	155	160
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<210> 19

<211> 4384

<212> DNA

<213> Homo sapiens

<220>

<223> Clone 14H12 - cDNA FLJ10838 fis, clone
 NT2RP4001274, weakly similar to Human transporter
 protein (gl7) mRNA

<220>

<221> CDS

<222> (534)..(1754)

<400> 19

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                                         1

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Lys Gln Asn Leu Ile Leu Leu Thr Phe Val Ser Ile Phe Ser Leu Tyr
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Ser Val His Leu Leu Leu Lys Thr Ala Asn Glu Gly Gly Ser Leu Leu
          20                      25                      30

tat gaa caa ttg gga tat aag gca ttt gga tta gtt gga aag ctt gca      680
Tyr Glu Gln Leu Gly Tyr Lys Ala Phe Gly Leu Val Gly Lys Leu Ala
          35                      40                      45

gca tct gga tca att aca atg cag aat att gga gct atg tca agc tac      728
Ala Ser Gly Ser Ile Thr Met Gln Asn Ile Gly Ala Met Ser Ser Tyr
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Asn Ile Glu Asp Lys Thr Gly Leu Trp Tyr Leu Asn Gly Asn Tyr Leu	
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Val Leu Leu Val Ser Leu Val Val Ile Leu Pro Leu Ser Leu Phe Arg	
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Asn Leu Gly Tyr Leu Gly Tyr Thr Ser Gly Leu Ser Leu Leu Cys Met	
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Val Phe Phe Leu Ile Val Val Ile Cys Lys Lys Phe Gln Val Pro Cys	
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Pro Val Glu Ala Ala Leu Ile Ile Asn Glu Thr Ile Asn Thr Thr Leu	
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Thr Gln Pro Thr Ala Leu Val Pro Ala Leu Ser Arg Asn Val Thr Glu	
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245 250 255	

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<210> 20

<211> 406

<212> PRT

<213> Homo sapiens

<223> Clone 14H12 - cDNA FLJ10838 fis, clone
NT2RP4001274, weakly similar to Human transporter
protein (g17) mRNA

<400> 20

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 85 90 95
 Leu Val Leu Leu Val Ser Leu Val Val Ile Leu Pro Leu Ser Leu Phe
 100 105 110
 Arg Asn Leu Gly Tyr Leu Gly Tyr Thr Ser Gly Leu Ser Leu Leu Cys
 115 120 125
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 130 135 140
 Cys Pro Val Glu Ala Ala Leu Ile Ile Asn Glu Thr Ile Asn Thr Thr
 145 150 155 160
 Leu Thr Gln Pro Thr Ala Leu Val Pro Ala Leu Ser Arg Asn Val Thr
 165 170 175
 Glu Asn Asp Ser Cys Arg Pro His Tyr Phe Ile Phe Asn Ser Gln Thr
 180 185 190
 Val Tyr Ala Val Pro Ile Leu Ile Phe Ser Phe Val Cys His Pro Ala
 195 200 205
 Val Leu Pro Ile Tyr Glu Glu Leu Lys Asp Arg Ser Arg Arg Arg Met
 210 215 220
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 275 280 285
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 Leu Pro Ser Ala Phe Tyr Ile Lys Leu Val Lys Lys Glu Pro Met Lys
 355 360 365
 Ser Val Gln Lys Ile Gly Ala Leu Phe Phe Leu Leu Ser Gly Val Leu
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 Ala Pro Gly Gly Gly His
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<210> 21
 <211> 588
 <212> DNA
 <213> Homo sapiens

<220>

<223> clone 1F2 - hypothetical protein HSPC014 (HSPC014)

<400> 21

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 <213> Homo sapiens

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<223> clone 2H4 - hypothetical protein MGC2577
 (MGC2577)

<400> 22

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accctaagt gaaaatgtta gtgaactaaa ggaaggagcc attcttgaa ctggacgact 480
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 <211> 533
 <212> DNA
 <213> Homo sapiens

<220>
 <223> clone 3C2 - clone FLC0593

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 <213> Homo sapiens

<220>
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 ttaaaat 547

<210> 25
 <211> 537
 <212> DNA
 <213> Homo sapiens

<220>
 <223> clone 4D10 - full length insert cDNA clone ZE03C06

<400> 25

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acaaaaaaaa tttatggccc aaaatgacca acgaaattgt tacaatagaa tttatccaat 420
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<210> 26

<211> 582

<212> DNA

<213> Homo sapiens

<220>

<223> clone 4E9 - hypothetical protein dJ465N24.2.1
(DJ465N24.2.1)

<400> 26

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<210> 27

<211> 388

<212> DNA

<213> Homo sapiens

<220>

<223> clone 4F7 - chromosome 1 clone RP11-109I2

<400> 27

```

tttttttncc ctggatcagc tcggcctnna aggcctcgga aaacaagatg cttcctctgg 60
aatgtgagtc caaagagtta ccagcgctgc cctctagtga totcagctca ggntatgcac 120
taaccgtgtg gntacagggc tgagtagtgc tgacgtgtga agtgaatgga aggcctcgag 180
gtgtttgtgg ctggccaccc tgatcagcct gcaggtagtc ccgatgaagc cagggcacag 240

```



```

ggggattcgt tccagcttgt tcactttatt ctgccttgcc aggttactga aagtcacctcg 300
tttgctctca ccagccttcc tggaaatgtg gactcttgaa agaaaagctc cegtgcctctt 360
gaagtatacc tgcttgccan gggagtc 388

```

<210> 28

<211> 605

<212> DNA

<213> Homo sapiens

<220>

<223> clone 4S-10d4 -DNA sequence from clone RP1-64K7 on chromosome 20q11.21-11.23 Contains the EIF2S2 gene for eukaryotic translation initiation factor 2 subunit 2 (beta, 38kD), a putative novel gene, the

<220>

<223> ribonucleoprotein RALY or autoantigen P542, an RPS2 (RPS4) (40S ribosomal protein S2) pseudogene, ESTs, STS, GSSs and two CpG islands

<400> 28

```

tgcttggtca tgctggcacc gggtcatatg ctggacaggg agaacgagag tcccatcctg 60
gaaatccaga aaagcccttg gatgctccag cccctgggaa agcacacagc caggcccttg 120
ggtgggaggt tggcttctaa cagtgcatac acatgccctt cctctgagtc ggggcagcaa 180
aaacatccat tccgctgcgc aacagttgtc atttttctaa catctgaaaa ctccagaagg 240
agatggtgat aaatgtggta ccggattctg cctaaaggat cagtcttttag atgttttcag 300
attgaaagcc tcatttgtga tcctcacagc catcttgaaa gaatagagca gccagtgagg 360
atactggatt gtgagctaag aggcttggga ctttccccct gttgctgcca gccaggttga 420
tgaccctggg caagtctttt tccttaccag gtctcagttt cctcagctgt aaaatgagag 480
gttgatctgg atcagggata gtaaattgggc ctttgttcag ttactgactg ttgtataaca 540
aaccaccccc aaatttagta gccttaataa acatttatta gtccttgaaa aaaaaaaaaa 600
aaaaa 605

```

<210> 29

<211> 661

<212> DNA

<213> Homo sapiens

<220>

<223> clone 4S-10G6 - similar to C50F4.16.p (LOC256281)

<400> 29

```

tgggggtgagt ggccaagact ggcctctgtc tagaaccctg gagtctcact ggagatccag 60
gttggggggcc acctggctga ggaaccatga gacaccaaag atgacgccga gggctcttggg 120
gatgtccggg tcgtctgcaa aggcctgggc gccttcagg ggcaggtgca ccacctcaat 180

```

```

gagctcacc ctcctccacca ggccccccacc tggaccgcta cgctgggcat ctgtcacctc 240
tgtgtagaac atgggtctgtc tggagccagt cagtcccact ccagaccagt atgtggcgac 300
ccggcgcgaga tcagagggggg ccaagtggta gccacactcc tcccaagcct ccttgcaagc 360
cacttcctcc agcgagagacc caggctggtc cacgaggccg gcacacagct caactgtcac 420
ccccgctgag ccgggcaggc ctggctgtag ctcccgaggc ccgtcctggc ctacagctgc 480
tagggaccct gggaagcggc gctccacctc acccgcatat acagctggcc ggaactgctt 540
caccaacacc aggtcctcctc gagaagagtt gaataagaga acggtcacgc tgtcatgcgt 600
cttcatgaag tcccaggact tctgggcacc attctggcgg taatgcagcg tgagcggccg 660
c

```

<210> 30

<211> 667

<212> DNA

<213> Homo sapiens

<220>

<223> clone 4S-11B8 - DNA sequence from clone RP11-735A5
on chromosome 1

<400> 30

```

tttttttttt tttttttaaa gaaaacctac tgtatgccat ttgcaagaga tataacaaaa 60
gcaaaactgat atgaaaaggc tgaaataaag ggatagttgg tttctggaaa aactaacaac 120
aacaacaaaa atcttccatg aatttttgtt gttgttattt taattttttt tgagacacgg 180
tcttgctgtc acccaggctg gagtacaggc gcacaatcat ggctcacggc agcatcgacc 240
tcctaggctc aaacgatcct ccacttttag cttcccaa at agctggaact acaggtgtgc 300
accgccacgc ctggctactt aaattttttt tttttttaat ttgtacagac atctcaccat 360
gttgcccaag ctggtctcca gctcctgggc tcaagtgatt caccacacct gcactctcaa 420
agtgtgaggc ttataggcat gagccactgt actcggcatt ccactgatac ttgataatga 480
aaaaaaagag tottaatacc caatattaga aatacaaaga gaaacagaag aacaaatcag 540
caaaggttta aaagaacaaa aattttatat atatatat ataaacaaat tgatctcaat 600
aaataacaaa ataaacaaaa tggacaaact cttaaaaaa tataccttac caaaactacc 660
tcaaaaaa

```

<210> 31

<211> 578

<212> DNA

<213> Homo sapiens

<220>

<223> clone 4S-11H3 - cDNA FLJ14279 fis

<400> 31

```

ttgtcaaaaa tccattcctg tcccctccac cctgttttta ttctctatc cactggaaac 60
cattaaaaaa aaaagttttt gatttatagt caattcttgt atttgaggta gttgtgttct 120
ataaagtgtg gcaaatactg aattagcaaa taccaaatta ttactactag ggcaaataca 180
cgattaagtt tctgtgagcc tctggtaata tttttgtcaa ccaatcgata catttacttt 240

```

gttttatgag tgtttctgtt aaagatacct tatttagtat atattgttga ttcattaacg 300
ttaaactcac tgccaatagt attaaaaactc atatgaatga aatgtgtcta acacgtattt 360
tccttataag tcatatcatg gccttcttga gcttaagaac actagagagc acttcagcac 420
tacacttgag ggccatttta tttacttatt tttacagatg gatcttgcta tattgcccac 480
actggactgt aatggctatt cacagggtgcc aaagtagcat actgcagact caaactcctg 540
gcctcaagcc atccacctca tcctcccaag tagttggg 578

<210> 32

<211> 352

<212> DNA

<213> Homo sapiens

<220>

<223> clone 4S-12A9 - chromosome 18, clone RP11-13N13

<400> 32

acatcacttg aggtcagtag acagggtttc accatgttac ccaggctggt ctcgaaactcc 60
tgacctcaag tgattcgccc atctcggcct gccaaagtgc tgaggtcaca ggcatgaggt 120
caggagaacg agaccatcct aacacagcga aaccccgctc ctactaaaaa gacaaaaaat 180
tagctgggtg tgggtggcagg cacctgtagt cccagctact cgggaggctg aggcaggaga 240
atggcctgaa cccgggaggc agagcttgca gtgagccaag atcgcgccac tgtactccag 300
cctgggcaac agagcaagac tgtctccaga aaaaaaaaaa aaaaaaaaaa aa 352

<210> 33

<211> 469

<212> DNA

<213> Homo sapiens

<220>

<223> clone 4S-12E6- hypothetical protein MGC955

<400> 33

gggagagtgg tcagcagtat ttttccatca tttctccaaa ggaatggggg acaagttgtc 60
cacatgactt ccttgggtgcc taaaaactac agcatgactt gtcctggact ccgtatgagg 120
acattgagaa gcaagatgct aaaatcagca tgatggacat gttgctaagc cagtcagtgg 180
ccctgcctcc gtgcactgaa cccaacttcc agggactgac tcaactgagag tgggctttga 240
caaacagctc tcacaggacc tggctgtcaa cctccttggt gccccactg ttgccttgag 300
aattgaagac atgtaggtga ctcaaaaact tcttggaaag agaccctgtg tgaatgtaaa 360
tgctgtcatt atgactttta attgggatgg gaataatcat tgagacagag tcaactgtctt 420
tcgggatcct ctttggacca cagataccca agtcagtcag tttcagagt 469

<210> 34

<211> 795

<212> DNA

<213> Homo sapiens

<220>

<223> clone 4S- 14D4 - clone IMAGE; 4778940 mRNA

<400> 34

```

aaaagcagat ttaaaaagca gctcacagtt gggcgcggtg gctcacacct gtaatcccag 60
cactttggga ggccgaggcg ggtggatcac cagagggtcag aagttcgaga ccagcctggc 120
caacgttggtg aaaccctgtc tctactaaaa atacaaaaat tggccacgtg tgggtggcggg 180
cacctgtaat cctagctact tgggaagctg agacaggaga attgcttgaa ccgaggaggt 240
ggagtgcagt ggcatgatta aggttcactg cagcctcaat ctcccactct ccagcgatca 300
tcccacctca gcctcttggg tagctgggac cacaggcacg agctaccatg cctggctaata 360
ttattttttg tagagacggg gtttcgccat gttgccccagg ctggtctgga atttctgacc 420
tctggcagtc cacctgtttc cgctcccaa atgctgagat tagaggcatg agtcactgca 480
cccagcccg cgcctctttt ataaggcat taatcccctt catgagggat attctctcat 540
gacttaata cctgccaag accccacctc ttaatactac attaagtgat gggttcaatg 600
tattnaattg gggggggagg gggcacattc agaccatagc atcttagtca ttcttggtt 660
tatttaagat tatttagact ganggcattg aaaaatagca tacttggatg ggacttcagc 720
attcnatnag tggccttaat aaaacctggt aattaaaaaa gcncttaatt ttgcaaaaaa 780
aaaaaaaaa aaaaaa

```

795

<210> 35

<211> 324

<212> DNA

<213> Homo sapiens

<220>

<223> clone 4S-2A6 - PAC clone RP3-414A15 from 14q24.3

<400> 35

```

tggcgtgaac ccagagggtg gagcttggtg gaggtgacag cgtgctggca gccctcacag 60
cacttctct gcctgggctc ccactttggc ggcacttgag aagcccttca gccaccgct 120
gcactgtggg agcccctttc tgggctggcc aaggccggag ccgtctccct cagcttgag 180
ggagggtgtg agggagaggc gcaagtggga actgaggctg cgttccgcac ttgcgggcca 240
gctggagtgc cagggtgggca tgggcttggg gggccccaca ctcgagagccg ccagctggcc 300
ctgccagccc cgggcaatga gggg

```

324

<210> 36

<211> 226

<212> DNA

<213> Homo sapiens

<220>

<223> clone 4S-2D3 - DNA sequence from clone RP11-357H24
on chromosome 10

<400> 36

```

gtctgctgcc tactcgtatg taatatgtgt acataaaagc ggcagctggg ttttcgttta 60
agagtaatct aatatacaga atttgggccc ttaagggttt atacctcttc atttaaaatg 120
ctttctggac aatctgctac caaacacatt ttgttatagg tgacattaaa actacatata 180
aatctacctg cagcacaaca cataaaaaaa aaaaaaaaaa aaaaaa 226

```

<210> 37

<211> 377

<212> DNA

<213> Homo sapiens

<220>

<223> clone 4S-2E2 - cDNA FLJ14385 fis, clone
HEMBA1002212, weakly similar to TYROSINE-PROTEIN
KINASE 2

<400> 37

```

gttggccttt gtttaaaaca ctgaaccttt tgctgatgtg tttatcaaata gataactgga 60
agctgaggag aatatgcctn aaaaagagta gtccttgga tacttcagac tctggttaca 120
gattgtcttg atctcttgga tctcctcaga tctttgggtt ttgctttaat ttattaaatg 180
tattttccat actgagttta aaatttatta atttgtaact taagcatttc ccagctgtgt 240
aaaaacaata aaactcaaata aggatgataa agaataaagg acactttggg taccagaaaa 300
aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa 360
aaaaaaaaaa aaaaaaa 377

```

<210> 38

<211> 758

<212> DNA

<213> Homo sapiens

<220>

<223> clone 4S-2F8 - chromosome 17, clone hRPC.1110_E_20

<400> 38

```

caaaaaaaaaa aaaaaaaaaa aaaaaacatc agatcttgtg agacttattc actatcagga 60
gaccagcaca cgaggaaagac cctcatgatt caattacctc ccaccagggt cctcccacaa 120
cacataggaa ttatgggagc tgcaattcaa gatgacaatt ggggtggggac acagcaaaac 180
cacatcacat gctgagctgt agcagggtgaa taaaccactg agactacgaa cctctgtcct 240
ctgaagaaga tgccattttc tcaaattcta ggagtgtggg ccaactgttct gaggttgctg 300
cagaggagca ctgctggaga aaaggaaggg ggaaatccac atatccacta agacatcgtc 360
agaatactgc acatggaatt ataggatatt aacaatttaa gagactgtag acacaggata 420
gtcaaaatct cccattttac agacaaatca acagaggccc agagcagtga aggcatctac 480
ccagacactc actggatcgg tgcagctgga tctagatcca ggtctcttga ctcatttaac 540
agctgttaaa ccaaaaatgg gtgtgattta gtcccattgt catctgatac attggcaatg 600
ccctgcataa ttttttgtct ctatgtttta tactcctttg taaagggtag cttttgatata 660
ttcctgaact gagcatctgt taaaattgga ttcactcttc cttaaggca acaattgggtg 720
tttctgatct ttaatgccaa aaaaaaaaaa aaagaaaa 758

```

<210> 39
<211> 840
<212> DNA
<213> Homo sapiens

<220>
<223> clone 4S-3C2 - clone FLC0593

<400> 39
tgactttacc gaactgacag ccattgggga ggcagatgcg ggtgtggagg tgtgggctga 60
aggtagtgac tgtttgattt taaaaagtgt gactgtcagt tgtatctgtt gctttttctca 120
atgattcagg gatacaaatg ggcttctctc attcattaaa agaaaacgcg acatctttct 180
aagattctct gtgggaaaat gactgtcaat aaaatgcggg tttctgggcc attcgtctta 240
ctttcatttt ttgattacaa atttctcttg acgcacacaa ttatgtctgc taatcctctt 300
cttcctagag agagaaaactg tgctccttca gtgttgctgc cataaagggg tttggggaat 360
cgattgtaaa agtcccaggt tctaaattaa ctaaagtgtg acagaaatga acgtgtaagt 420
aatgtttcta caggctcttg caacaaactg tcactttcgt ctccagcaga gggagctgta 480
ggaatagtgc ttccagatgt ggtctcccggt gtggggccca gcaatggggg cccctgatgc 540
caagagctct ggaggttctt gaaagagggg acacgaagga ggagtgactg ggaagcctcc 600
catgccaaagg aggtgggagg tgccctggaa atagctgcct catgccactt aggccatgac 660
tggttttaat gtcagtgggt tgccacagtg cagtggctag acaactgaaa ggggctacca 720
aggctgggaa aaaaatgcaa ttgttgctgt gagtgacttt gaaagactct ggtgccttgt 780
ggtgcccttc tgaaattcaa acagtaatgc aaaagtgtct gcattagaat ttacgggtgc 840

<210> 40
<211> 480
<212> DNA
<213> Homo sapiens

<220>
<223> clone 4S-3F11 - BAC clone CTD-2314H8

<400> 40
agagttgctg aaaaatggag taacagtgtg tgggaacttca catcagttct ccttattgat 60
tgttagtttg atocctcttg ttcttttgtt gtaaacattt tctataatta ggaaatgcc 120
tttaagagtg agagaggtat atatctatga gccatttgtt ttggtgtttt acaagaactt 180
taccatactg gtgtgtagtc cattctgtac agttttaaag tgattcacga tttgcaggct 240
ttttatcaga tcacaaaaaa atcagtcctt aagcatttgc ttggttaagg ttcttaagat 300
taggtttata atacaacat ctgtaatgta tctctcgttt gagcttgtgg gccatacaat 360
tcattaacta gatgaataca ttgtggacag catcctcact acccctctct actcactcac 420
aaagaacat gatacactgg aatgtttttc tctggaatcc tctttctact cttgtattaa 480

<210> 41
<211> 506

<212> DNA

<213> Homo sapiens

<220>

<223> clone 4S-4C11 - similar to putative, clone
MGC:33177 IMAGE:4823662

<400> 41

```
atcatttgat aatttacctt agagcattta aaaaaatata atcaaactaa ttgccagcca 60
agtcagtcac cctcctggga gtatatagag tcccaagggt agcgctcctg tattagacta 120
tttcaatttt aggaaaatca tgaccatgtg gggaaacaat gactttaaaa tgctgaaatt 180
aaaatttatg ctttaactgg aatatttttt gcttaactac tcaattagaa tattgtacac 240
ctgatcaatg tgtgttcagc acagatggcc atgaattgtc atttatagtc caatttttta 300
tcttaatcat aaaatgttta ggaatctatg aaatttaact ttaggaacaa aacgttttagc 360
agggttgatt gatattatgt ttacattgtt ctggcaatcc acagaaagag aagagcctta 420
atttttaaaa cccatttttag tcattttatg acaattaaag ttgtttaata aacatctttt 480
ttcaaagaag caaaaaaaaa aaaaaa                                     506
```

<210> 42

<211> 558

<212> DNA

<213> Homo sapiens

<220>

<223> clone 4S-4E9 - hypothetical protein dJ465N24.2.1
(DJ465N24.2.1)

<400> 42

```
ctgtcggaaa aggtaacaga agatggaact cgaaatccca atggaaaacc taccagcaa 60
agaagcatag cttttagctc taataattct gtagcaaagc caatacaaaa atcagctaaa 120
gctgccacag aagaggcatc ttcaagatca caaaaaatag atcagaaaaa aagtccatat 180
ggactgtgga tacctatcta aaagaagaaa actgatggct aagtttgcat gaaaactgca 240
ctttattgca agttagtgtt tctagcatta tcccatccct ttgagccatt caggggtact 300
tgtgcattta aaaaccaaca caaaaagatg taaatactta aactcaaat attaacattt 360
taggtttctc ttgcagatat gagagatagc acagatggac caaagggttat gcacaggtgg 420
gagtcttttg tatatagttg taaatattgt cttgggttatg taaaaatgaa atttttttaga 480
cacagtaatt gaactgtatt cctgttttgt atatttaata aatttcttgt ttccattctt 540
aaaaaaaaaa aaaaaaaaaa                                     558
```

<210> 43

<211> 352

<212> DNA

<213> Homo sapiens

<220>

<223> clone 4S-4F7 - chromosome 1 clone RP11-109I2

<400> 43

```

gggaaaacaa gatgcttcct ctggaatgtg agtccaaaga gttaccagcg ctgccctcta 60
gtgatctcag ctcagcatat gcactaaccg tgtgtttaca gggctgagta gtgctgcagt 120
gtgaagtgaa tggaaggcct cgaggtgttt gtggctggcc accctgatca gcctgcaggt 180
agtcccgatg aagccagggc acagggggat tctgtccagc ttgttcactt tattctgcct 240
tgccagggtta ctgaaagtcc ctctgtttgct ctcaccagcc ttcttggaat tgtggactct 300
tgaaagaaaa gctcccgctg tcttgaagta tacctgcttg ccaggggagt cc 352

```

<210> 44

<211> 524

<212> DNA

<213> Homo sapiens

<220>

<223> clone 4S-5C4 - similar to KIAA0674 protein

<400> 44

```

cccttcctta agcatgattt tgcacagcca accctgggtc taggcgaacc acaggggtgag 60
gtcaagggtga gcattctggg aacaatattt gggctcagag ggtgggttgg ccaccttctg 120
agccccaccc cgcagcagacc tggatgaagag gatcataacc ctgtcttcaa gaacactggg 180
atttcagcag caagttggaa gaaggactgg taggttcccc tccaagccag tcacctgtaa 240
gagtctgtgc ctctgccaga ctttttaatc tcttcattaa ctctcagact gacctgggag 300
ccctcctcta cctgaatcca gtgctcaact gtgccccggc aacaagacct gggctgaggt 360
ctccctggta gaactaaggg agattacacc atctaaatcc cagtgcagtc aacagcctgg 420
cctatagtcc tgggacatgt atcttcttct ttgccttaaa tctgatacaa gaggtcaatg 480
actttgaaaa taaaactaaa ataaatgtca aaaaaaaaaa aaaa 524

```

<210> 45

<211> 891

<212> DNA

<213> Homo sapiens

<220>

<223> clone 4S-6F6 - similar to RIKEN cDNA 1110012M11 gene

<400> 45

```

atggattcca aggcctctgt gtccaagaag aaacgcatgt gtgtgaagct gttgcccttg 60
ggagccacgg acacggctgt gtttgatgtc cggctgagtg ggaagaccaa gacagtgcct 120
ggataccttc gaatagggga catgggcggc ttgcatctt ggtgcaagaa ggccaaggcc 180
ccgaggccag tgcccaagcc ccgaggtctc agccgggaca tgcagggcct ctctctggat 240
gcagccagcc agccaagtaa gggcggcctc ctggagcggg cagcgtcaag gctgggctct 300
cgggcatcca ctctgaggag gaatgactcc atctacgagg cctccagcct ctatggcatc 360
tcagccatgg atggggttcc cttcacactc caccacgat ttgagggcaa gagctgcagc 420
cccctggcct tctctgcttt tggggacctg accatcaagt ctctggcgga cattgaggag 480

```



```

gagtataact acggcttcgt ggtggagaag accgcggctg cccgcctgcc cccagcgtc 540
tcatagtccc tcacccttcc gcggaaagag ccccttact ccacctcccc gccagcctgg 600
ggccaccccc cctcactgca tcctgggaac cttcgccctg caaggcgttt gctatcttca 660
gccactgggc ggagctgcag ccctggagga gggggcgggt cgaggctgcg tggatgatgg 720
gtctccgccc ccacgccctg ccgggcaggg ctggagctgg acagaagcca gtgcctttaa 780
gtcatttgtg tcaaaaccct ctgggggtccg gaggtgtgtc ggggtgtcctc ctggcaataa 840
acactaccgg gttctcgcca aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa a
891

```

<210> 46

<211> 902

<212> DNA

<213> Homo sapiens

<220>

<223> clone 4S-6H8 - cDNA FLJ31039 fis, clone
HSYRA2000221

<400> 46

```

agagattctc taaatatgga attagattag gattctctgc tccacttaac acacattttt 60
aaattagtag tgatgattga gggatggaca atagcacacc aaaaaaaaaa agtttagtat 120
gaaaaattta aacctgttgg ttaagtcatt cccattaatg tcattttgct gagggtgact 180
tggtcctttt gaattgcttt ggtgtacggg tatgttctga tttttcatgc aagctcctct 240
gccattccac cgctctgagg agtaattgta gcacttcaca tgtgctgtgg ttgtgatcac 300
atggtgacat acatagcatg tgtgttccca gctgttgtgt gtttatgtga catttgatgc 360
caatacatat gtcttcaagg tatgcttgtt ccctcccagc togtggaata tcaaaaaaat 420
tcattgctgg aaaaattatt tcatagacaa aaatgttaat gttctcttgg ggacttagag 480
ttgaaaatat ttgtatagat ttggttctca agtccacaga atcgatatct ctgtgggtctc 540
cctttgggtg tcatctggga gccatgtgta tggaagattc tgtcacaggc ggctgggatg 600
tgggcagatg ctgttagcct ccctctccac gtgggtgtcc atgcctgacg tgtcccctag 660
ttcaaggaag cgccatcttt agcatgaaaa caattgcgtt cccctaggaa atgaagaaaa 720
aatgagctga aatttcctta tacatttgaa tttgttcatt ttttaaagag acatttttgt 780
tgtctgcttt gtggtactta taaaatttgt tttccattga aattgccatt tataaatttg 840
cagatatgta ctaatttaga ttttttttaa gtgttcaata aaataaggat atatttactg 900
tg
902

```

<210> 47

<211> 566

<212> DNA

<213> Homo sapiens

<220>

<223> clone 4S-7F6 - cDNA FLJ13305 fis

<400> 47

```

aagactctct tagtgactct tccagatctg tatcagaaaa gaactcctat caccctgtct 60
cattaatgac atcattttca gagcctgatt taggcagtc ttcctccttg tatgtgtcct 120

```

```

cctctgaaga ggagttaccc aacctagaaa aagagtatcc taggaaaaac agaattgatga 180
cctatgctaa ggagctcatc aacaatatgt ggacagactt ttgtgttgag gattatattc 240
gctgtaaaga tactggcttc catgcagctg aaaaaagaag gaagaaacga aaagaatggg 300
tgcccacaat tacagtaccg gagccttttc aaatgatgat aagagaactg aagaaaatct 360
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566

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<220>

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<223> clone 4S-9F4 - similar to hypothetical protein
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1021

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<212> DNA

<213> Homo sapiens

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<223> clone 5S-21/57 - hypothetical protein FLJ10081

<400> 51

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<400> 52

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 aaggagtccc agtatggatt taccaggc aatggacaga tgcccagggg cttggaagaa 420
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<400> 53

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taacaggaga gcagcaatac gattttgcca atggaataac agatttgccg gcattcactg 300
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<210> 54

<211> 23

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<220>

<223> Description of Artificial Sequence: primer used
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<211> 32

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<213> Artificial Sequence

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<223> Description of Artificial Sequence: primer used
for generation of a ferritin PCR fragment

<400> 55

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32

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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Organization
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PCT

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(71) Applicant (*for all designated States except US*):
**VLAAMS INTERUNIVERSITAIR INSTITUUT
VOOR BIOTECHNOLOGIE VZW** [BE/BE]; Rijviss-
chestraat 120, B-9052 Zwijnaarde (BE).

(72) Inventors; and

(75) Inventors/Applicants (*for US only*): **CONTRERAS,**
Roland, Henry [BE/BE]; Molenstraat 53, B-9820 Merel-
beke (BE). **CHEN, Cuiying** [BE/BE]; Lijsterstraat 10,
B-9860 Balegem (BE).

Published:

— with international search report

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4 March 2004

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.



WO 2003/045988 A3

(54) Title: METHOD TO ISOLATE GENES INVOLVED IN AGING

(57) Abstract: The present invention relates to a method to isolate genes involved in aging and/or oxidative stress, by mutation or transformation of a yeast cell, subsequent screening of the mutant or transformed cells that are affected in aging and isolation of the affected gene or genes, and the use of these genes to modulate aging and aging-associated diseases in a eukaryotic cell and/or organism.

INTERNATIONAL SEARCH REPORT

International Application No
PCT/EP 02/13549

A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 C07K14/39 C12N15/10

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C12N C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, EMBL, BIOSIS, MEDLINE

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 01 21786 A (KEIO UNIVERSITY ;NISHIMOTO IKUO (JP)) 29 March 2001 (2001-03-29) see example 2, p. 37, example 4, p. 40, example 6, p. 41 ----- -/--	8,13

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

* Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search

12 March 2003

Date of mailing of the international search report

24. 06. 03

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2
NL - 2280 HV Rijswijk
Tel. (+31-70) 340-2040, Tx. 31 651 epo nl,
Fax: (+31-70) 340-3016

Authorized officer

Paresce, D

INTERNATIONAL SEARCH REPORT

International Application No

PCT/EP 02/13549

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	MAX INGMAN, HENRIK KAESMANN, SVANTE PÄÄBO & ULF GYLLENSTEN: "Mitochondrial genome variation and the origin of modern humans" NATURE, vol. 408, no. 6813, 7 December 2000 (2000-12-07), pages 708-713, XP002190961 see abstract	8,13
X	-& DATABASE EMBL 'Online! 14 March 2001 (2001-03-14) "Homo sapiens mitochondrion, complete genome" Database accession no. AF346979 XP002234371 abstract	8,13
Y	--- HASHIMOTO Y., ET AL.: "A rescue factor abolishing neuronal cell death by a wide spectrum of familial Alzheimer's disease genes and A beta" PROC. NATL. ACAD. SCI. U.S.A., vol. 98, no. 11, 22 May 2001 (2001-05-22), pages 6336-6341, XP002190962 page 6336	8,13
X	-& DATABASE EMBL 'Online! "Homo sapiens Humanin (HN1) mRNA" Database accession no. AY029066 XP002234372 abstract	8,13
Y	--- US 2001/026930 A1 (AUSTRIACO NICANOR ET AL) 4 October 2001 (2001-10-04) see p. 9, right column, claims 1, 6	1-6,8,9, 13
Y	--- US 5 919 618 A (KENNEDY BRIAN ET AL) 6 July 1999 (1999-07-06) see columns 4-5, 16-17 and claims	1-6,8,9, 13
Y	--- WO 95 05459 A (MASSACHUSETTS INST TECHNOLOGY ;GUARENTE LEONARD P (US); AUSTRIACO) 23 February 1995 (1995-02-23) cited in the application see p. 28-33, claims 2, 10, 18, 26-33 --- -/--	1-6,8,9, 13

INTERNATIONAL SEARCH REPORT

International Application No

PCT/EP 02/13549

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	<p>GRZELAK A ET AL: "Decreased antioxidant defense during replicative aging of the yeast <i>Saccharomyces cerevisiae</i> studied using the 'baby machine' method"</p> <p>FEBS LETTERS, ELSEVIER SCIENCE PUBLISHERS, AMSTERDAM, NL,</p> <p>vol. 492, no. 1-2,</p> <p>9 March 2001 (2001-03-09), pages 123-126, XP004257348</p> <p>ISSN: 0014-5793</p> <p>see materials and methods and p. 124, left column</p> <p>-----</p>	<p>1-6,8,9, 13</p>

INTERNATIONAL SEARCH REPORT

International application No.
PCT/EP 02/13549

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
Although claims 9-10, 13-15 are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.
2. ☒ Claims Nos.: 7, 11-12
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
see FURTHER INFORMATION sheet PCT/ISA/210
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

see additional sheet

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☒ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
1-6 completely, 8-9, 13 partially

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
☐ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

Invention 1: claims 1-6 completely, claims 8-9,
13 partially

Claims 1-6 completely, claims 8-9, 13 partially are directed to a method to screen genes involved in aging, as well as genes isolated by said method, in particular a gene comprising SEQ ID NO: 1, the polypeptide expressed from said gene, and uses thereof.

Invention 2: claims 8-9, 13 partially

Claims 8-9, 13 partially are directed to a gene comprising SEQ ID NO: 3, the polypeptide expressed from said gene, and uses thereof.

Invention 3-45: claims 8-10, 13-15 partially

Inventions 3-45 are directed to genes or gene fragments involved in aging or oxidative stress. Each claimed sequence is considered a separate invention, the inventions are numbered according to the list of sequences given in claim 8, namely, invention 3 is directed to the gene comprising SEQ ID NO: 5, invention 4 is directed to the gene comprising SEQ ID NO: 7, etc. Note: claim 10 is searched partially for invention no: 7 and 10, claim 14 is searched for invention no: 7 and claim 15 is searched for invention no: 10. The order of inventions follows the order of sequences given for claim 8. The inventions are also directed to the polypeptides expressed from said genes, and uses thereof.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Continuation of Box I.2

Claims Nos.: 7, 11-12

Present claims 7, 11-12 and claim 9 partially relate to an extremely large number of possible genes or gene fragments. The claimed genes are characterized only by the fact that they may be isolated by a certain method. However, support within the meaning of Article 6 PCT and/or disclosure within the meaning of Article 5 PCT is to be found, however, for only a very small proportion of the compounds/products claimed. In the present case, the claims so lack support, and the application so lacks disclosure, that a meaningful search over the whole of the claimed scope is impossible. Consequently, the search has been carried out for those claims which appear to be supported and disclosed, namely those that refer to specific sequences or methods (claims 1-6, 8, 10, 13-15 and claim 9 partially).

The applicant's attention is drawn to the fact that claims, or parts of claims, relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/EP 02/13549

Patent document cited in search report		Publication date	Patent family member(s)	Publication date
WO 0121786	A	29-03-2001	AU 7313800 A	24-04-2001
			AU 7313900 A	24-04-2001
			CA 2385444 A1	29-03-2001
			EP 1221480 A1	10-07-2002
			WO 0121786 A1	29-03-2001
			WO 0121787 A1	29-03-2001
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US 2001026930	A1	04-10-2001	US 6218512 B1	17-04-2001
			US 5919618 A	06-07-1999
			US 5874210 A	23-02-1999
			WO 9505459 A1	23-02-1995
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US 5919618	A	06-07-1999	US 6218512 B1	17-04-2001
			US 2001026930 A1	04-10-2001
			US 5874210 A	23-02-1999
			WO 9505459 A1	23-02-1995
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			US 6218512 B1	17-04-2001
			US 5919618 A	06-07-1999
			US 2001026930 A1	04-10-2001
			US 5874210 A	23-02-1999
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